

Supplementary information for

## **Self-reinoculation with fecal flora changes microbiota density and composition leading to an altered bile-acid profile in the mouse small intestine**

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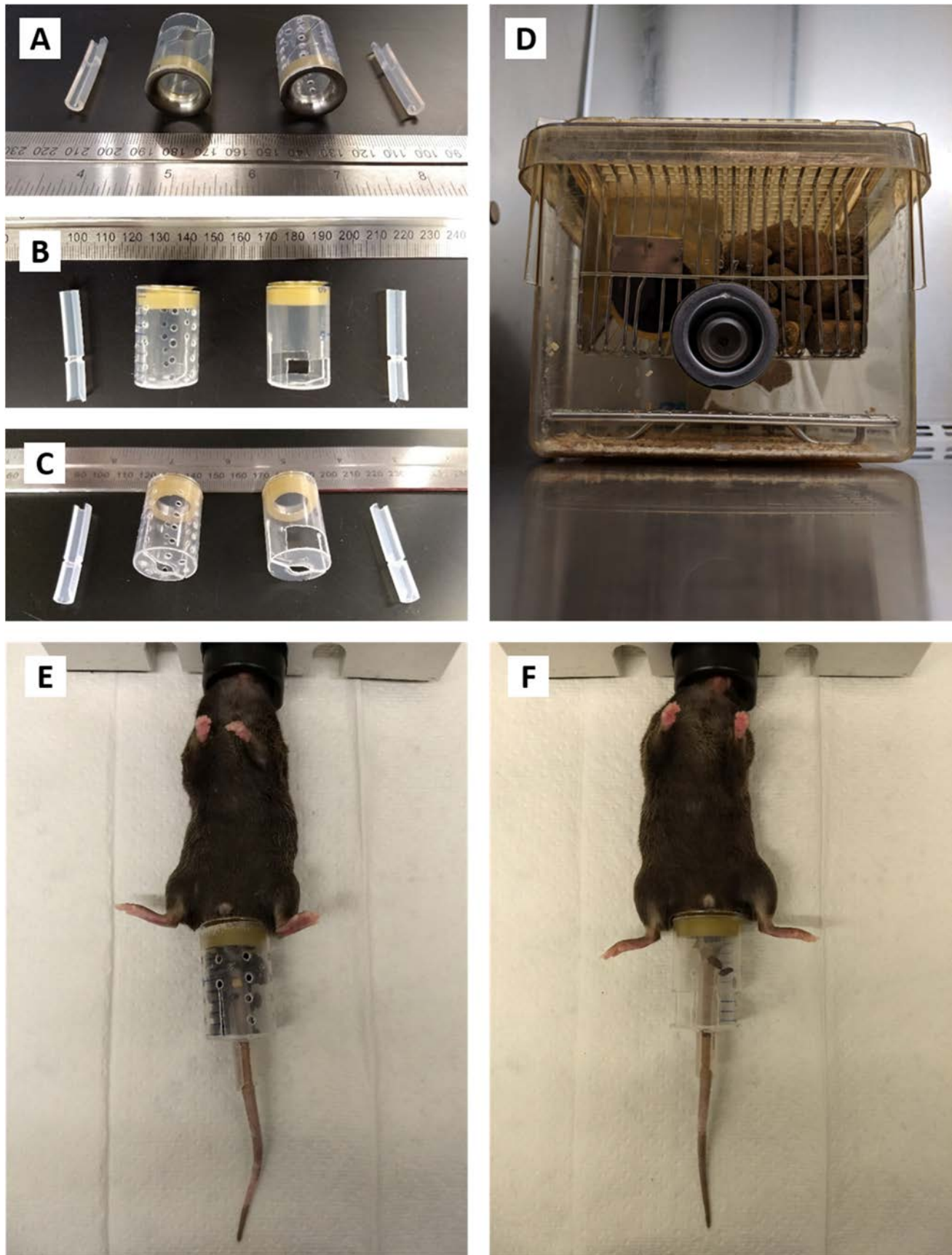
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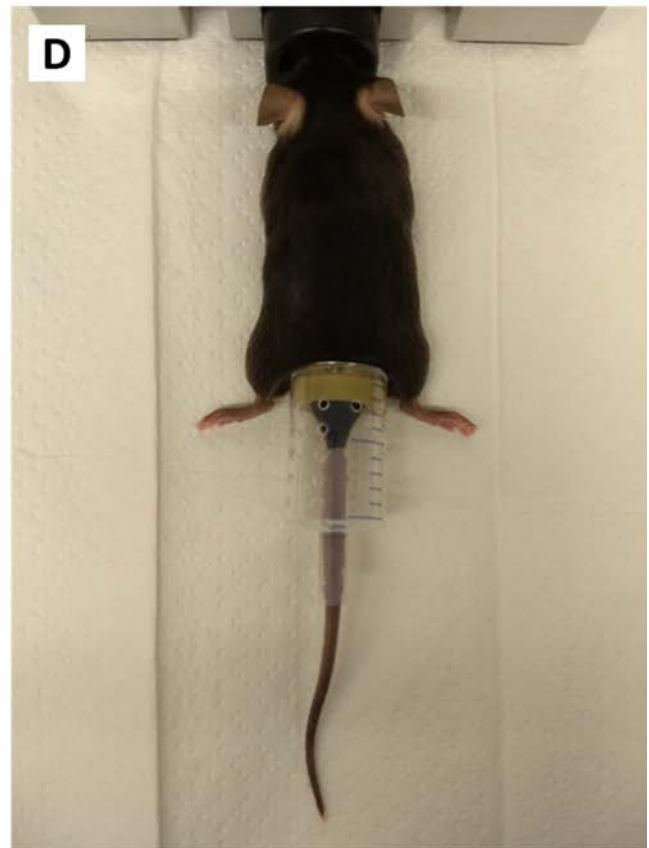
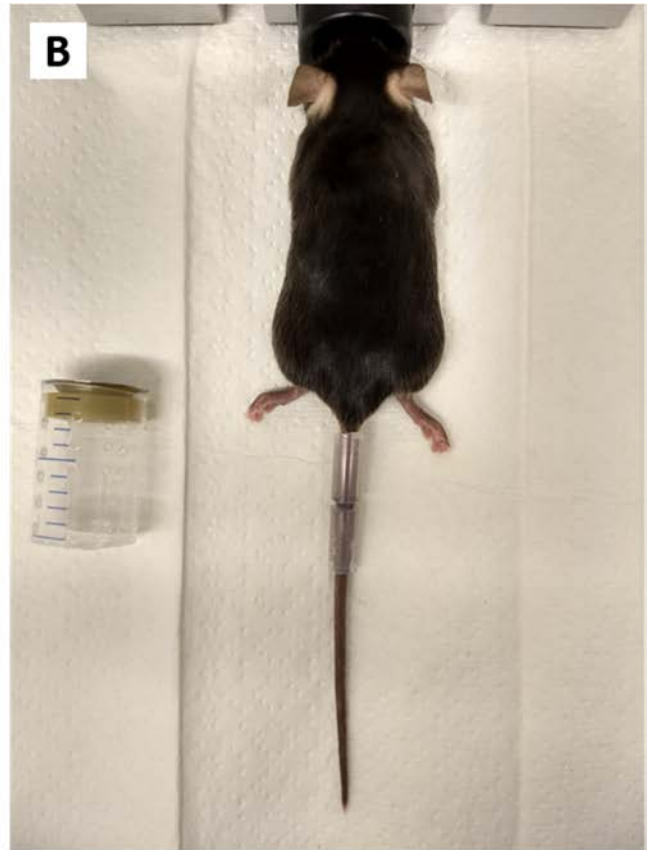
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**Figures S1-S5**

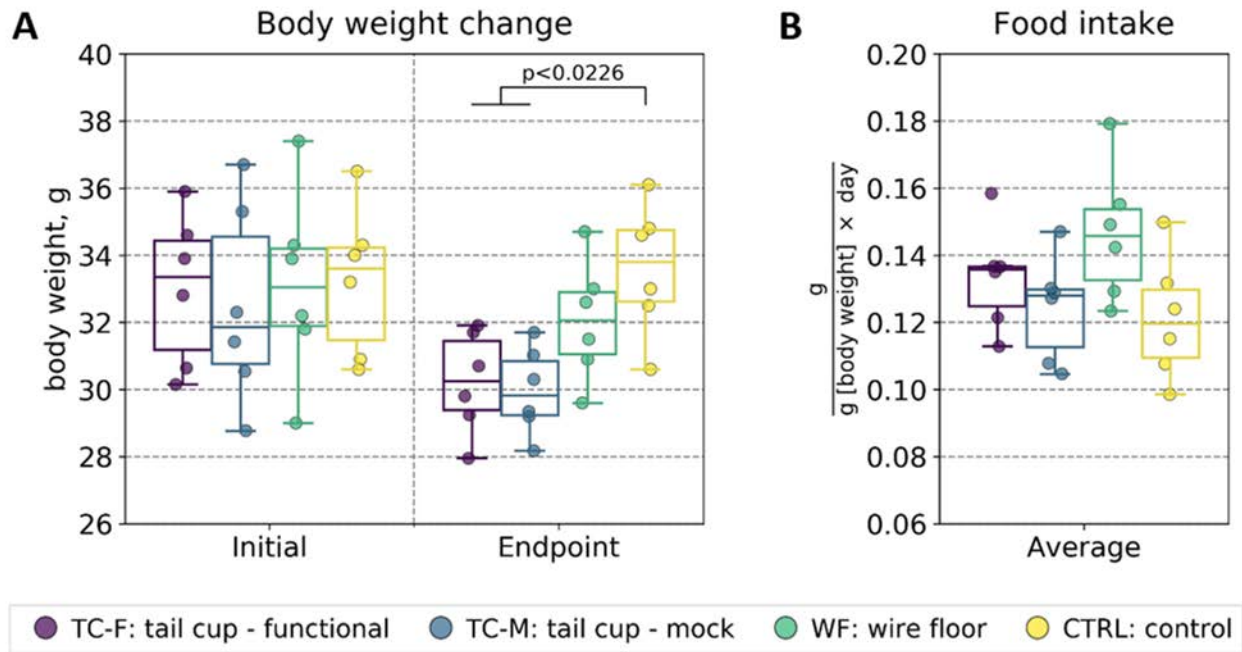
**Tables S1-S7**



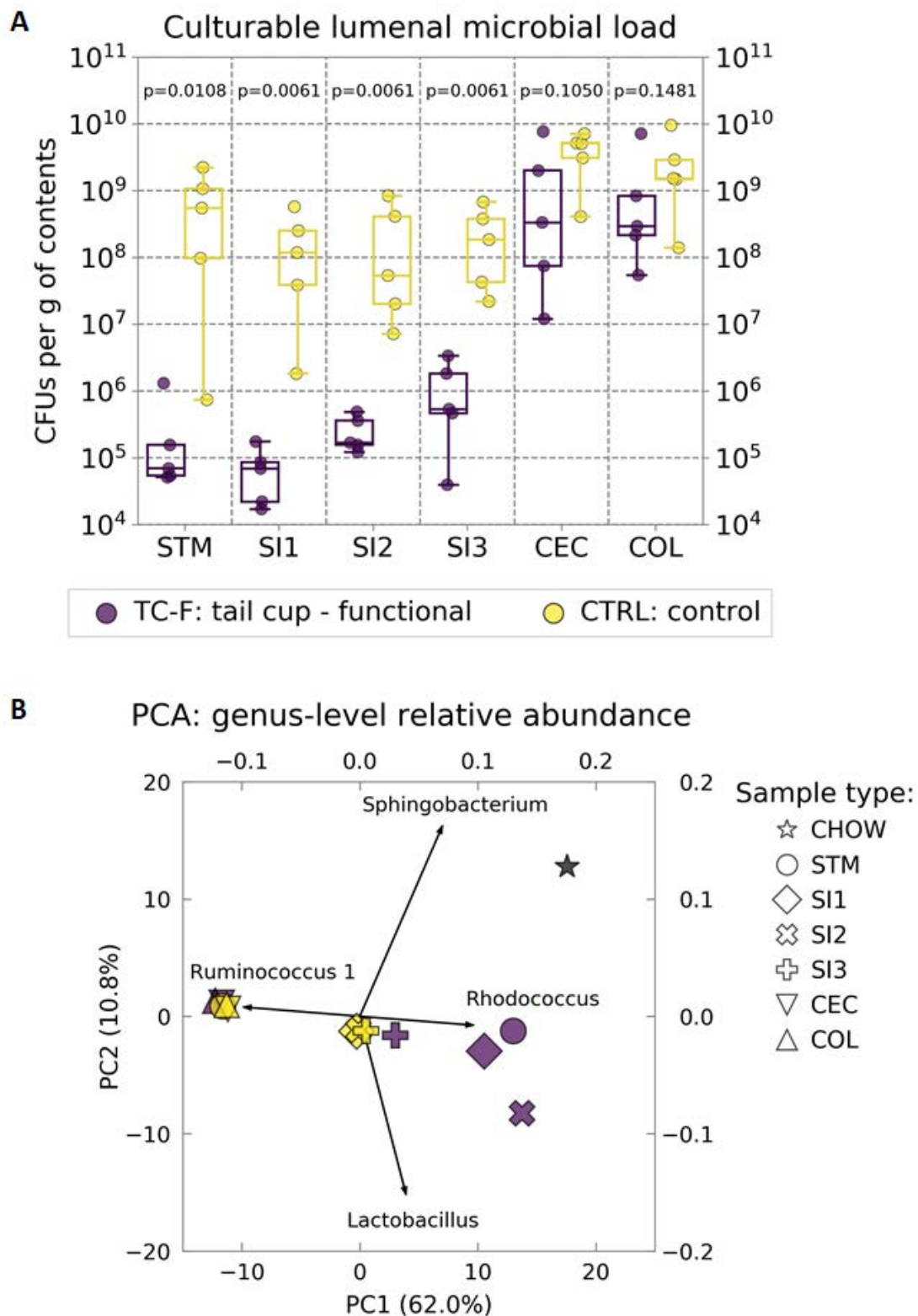
**Fig. S1. Tail cup design and experimental setup for preventing coprophagy.** (A, B, C) Functional (TC-F, left) and mock (TC-M, right) tail cups as viewed from different perspectives. (D) The standard cages with wire mesh floors used in this study (WF). (E, F) Ventral view of the functional (TC-F; left) and mock (TC-M, right) tail cups 24 hours after emptying (TC-F) or mock emptying (TC-M).



**Fig. S2. Mounting of functional tail cups onto mice. (A, B)** Ventral and dorsal view of the tail sleeve mounted at the tail base. **(C, D)** Ventral and dorsal view of the functional tail cup installed and locked in place using the tail sleeve.

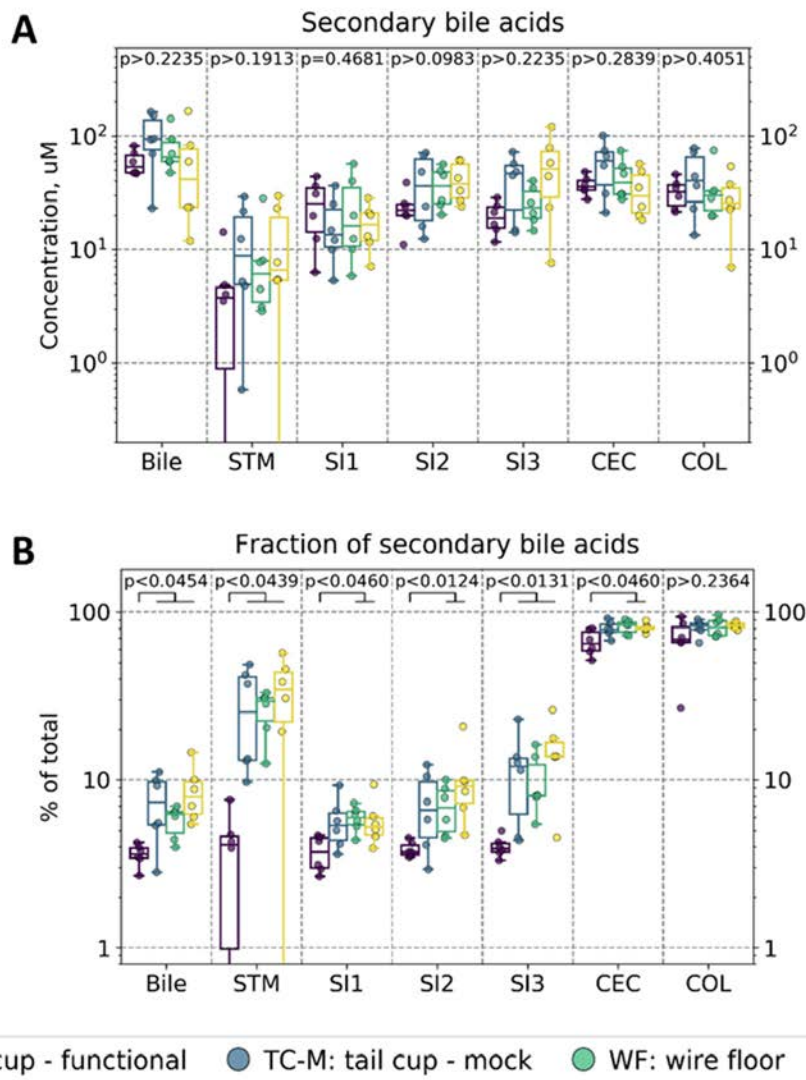


**Fig. S3. Body weight changes across all groups of mice in relation to food intake over the course of the study. (A)** Body weights of each individual animal at the beginning and at the endpoint of the study. **(B)** Normalized food intake per gram of body weight per day measured over the entire duration of the study. Multiple comparisons of the normally-distributed homoscedastic data were performed using one-way ANOVA; pairwise comparisons were performed using the Student's *t*-test with FDR correction. N = 6 mice per group.



**Fig. S4. Quantification of the culturable microbial load and microbiota profile along the entire GIT of mice fitted with functional tail cups (TC-F) and control mice (CTRL).** (A) Culturable microbial loads in contents along the gastrointestinal tract were evaluated using the most probable number (MPN) assay performed in anaerobic BHI-S broth (N = 5 mice per group, *P*-values were calculated using the Wilcoxon–Mann–Whitney test). (B) PCA analysis of the CLR-transformed relative microbial abundance profiles (16S rRNA gene amplicon sequencing) along the entire GIT in TC and CT mice (N = 1 mouse from each group).





**Fig. S5. Bile acid profiles in gallbladder bile and in luminal contents along the entire GIT.** (A) Total secondary bile acid levels (conjugated and unconjugated) and (B) the fraction of secondary bile acids (conjugated + unconjugated) in gallbladder bile and throughout the GIT (STM = stomach; SI1 = upper third of the small intestine (SI), SI2 = middle third of the SI, SI3 = lower third of the SI roughly corresponding to the duodenum, jejunum, and ileum respectively; CEC = cecum; COL = colon). In all plots, individual data points are overlaid onto box-and-whisker plots; whiskers extend from the quartiles (Q2 and Q3) to the last data point within  $1.5 \times$  interquartile range (IQR). Multiple comparisons were performed using the Kruskal–Wallis test; pairwise comparisons were performed using the Wilcoxon–Mann–Whitney test with FDR correction.  $N = 6$  mice per group.

**Table S1. Primer oligonucleotide sequences used in the study.** [NNNNNNNNNNNN] – 12-base barcode sequences “806rcbc” according to [148].

Primer	Oligonucleotide sequence	Assay	Reference
UN00F2	CAGCMGCCGCGGTAA	16S rRNA gene DNA qPCR 16S rRNA gene DNA ddPCR	[38]
UN00R0	GGACTACHVGGGTWTCTAAT		[147, 148]
UN00F2_BC	AATGATACGGCGACCACCGA GATCTACACTATGGTAATTGT CAGCMGCCGCGGTAA	16S rRNA gene DNA amplicon barcoding	[38]
UN00R0_BC	CAAGCAGAAGACGGCATAACGAGAT [NNNNNNNNNNNN] AGTCAGTCAGCC GGACTACHVGGGTWTCTAAT		[147, 148]
ILM00F(P5)	AATGATACGGCGACCACCGA	Barcoded amplicon and NGS library quantification ddPCR	[147–151]
ILM00R(P7)	CAAGCAGAAGACGGCATAACGA		
Seq_UN00F2_Read_1	TATGGTAATTGTCAGCMGCCGCGGTAA	MiSeq read 1	[38]
Seq_UN00R0_Read_2	AGTCAGTCAGCCGGACTACHVGGGTWTCTAAT	MiSeq read 2	[147, 148]
Seq_UN00R0_RC_Index	ATTAGAWACCCBDGTAGTCCGGCTGACTGACT	MiSeq index read	[147, 148]

**Table S2. Thermocycling parameters for the quantitative PCR (qPCR) assay for 16S rRNA gene DNA copy quantification.**

Step	Repeats	Temperature, °C	Time, sec
Initial denaturation	× 1	95	120
Cycle	× 40	95	15
		53-54	10
		68	45

**Table S3. Thermocycling parameters for the digital PCR (dPCR) assay for absolute 16S rRNA gene DNA copy quantification.**

Step	Repeats	Temperature, °C	Time, sec	Ramp, °C/sec
Initial denaturation	× 1	95	300	2.0
Cycle	× 40	95	30	2.0
		52	30	2.0
		68	60	2.0
Dye stabilization	× 1	4	300	2.0
		90	300	2.0
		12	∞	2.0

**Table S4. Thermocycling parameters for the 16S rRNA gene DNA amplicon barcoding PCR reaction for next generation sequencing (NGS).**

Step	Repeats	Temperature, °C	Time, sec
Initial denaturation	× 1	94	180
Cycle	× var.	94	45
		54	60
		72	105
Final extension	× 1	72	600

**Table S5. Thermocycling parameters for the digital PCR (dPCR) assay for barcoded amplicon and Illumina NGS library quantification.**

Step	Repeats	Temperature, °C	Time, sec	Ramp, °C/sec
Initial denaturation	× 1	95	300	2.0
Cycle	× 40	95	30	2.0



		60	90	2.0
Dye stabilization	× 1	4	300	2.0
		90	300	2.0
		12	∞	2.0

**Table S6. Reagents and chemical standards used in the bile acid metabolomics assay.**

<b>Bile acid</b>	<b>Reference #</b>	<b>Vendor</b>	<b>LOT</b>
TαMCA	C1893-000	Steraloids	B1439
TβMCA	C1899-000	Steraloids	B1594
TωMCA	C1889-000	Steraloids	B1731
THCA	C1887-000	Steraloids	B1621
αMCA	C1890-000	Steraloids	B1529
βMCA	C1895-000	Steraloids	B1725
ωMCA	C1888-000	Steraloids	B1710
HCA (gMCA)	C1850-000	Steraloids	B0696
HDCA	C0860-000	Steraloids	B0684
MCA	C0910-000	Steraloids	B1711
GDCA	C1087-000	Steraloids	B2122
GCA	C1927-000	Steraloids	
GHDCA	C0865-000	Steraloids	B1667
GHCA	C1860-000	Steraloids	L1105
TCA	13232UNL	Isosciences	EH1-2015-111A1
CA	13098UNL	Isosciences	EH1-2014-075A1
DCA	13100UNL	Isosciences	EH1-2014-076A1
TCDC	13105UNL	Isosciences	EH1-2015-110A1
TDCA	13225UNL	Isosciences	EH1-2015-112A1
TUDCA	13106UNL	Isosciences	EH1-2014-027A1
TLCA	13230UNL	Isosciences	EH1-2014-077A1
CDCA	13101UNL	Isosciences	PG1-2014-149A1
UDCA	13102UNL	Isosciences	EH1-2015-113A1
LCA	13099UNL	Isosciences	EH1-2014-030A1
D4-TCA	13232	Isosciences	SJ5-2015-035A1
D4-DCA	13100	Isosciences	RS6-2014-168A1
D4-CA	13098	Isosciences	SJ5-2015-100A1
D4-TDCA	13225	Isosciences	SJ5-2015-034A1
D4-GLCA	13231	Isosciences	SR3-2015-203A1
D4-GUDCA	13224	Isosciences	SJ5-2017-206A1

D4-GCDCA	13104	Isosciences	SJ4-2012-070A1
D4-GCA	13443	Isosciences	SJ5-2015-118A1
D4-GDCA	13226	Isosciences	SJ5-2015-033A1

**Legend for Table S7 (attached as .csv file):**

**Table S7. Bile acid concentrations in gallbladder bile and in luminal contents along the entire GIT.**