

Gene-targeted microfluidic cultivation validated by isolation of a gut bacterium listed in Human Microbiome Project's Most Wanted taxa

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Supporting Text

All chemicals were purchased from commercial sources and used as received unless otherwise stated.

Homemade Reagents

A protocol for making GBSS buffer can be found on the Schmidt Lab website (1). For H₂Oc or PBSc, 0.2% cysteine was added and the solution was steriley filtered through 0.22 µm membrane. 0.1 M Fe(SCN)₃ solution was steriley filtered through 0.22 µm membrane (Fisher Scientific) and used as a red dye solution. Tetradecane (Fisher Scientific) was steriley filtered through 0.22 µm membrane (Fisher Scientific). For assembling SlipChip for depositing PCR reagents, filtered tetradecane was degassed under house vacuum overnight. All plastic consumables and reagents were equilibrated in an anaerobic chamber for more than 24 hours before usage.

Anaerobic Chamber

A Coy lab anaerobic chamber equipped with dehumidifier was used in all anaerobic cultivation experiments. The hydrogen level was maintained at 3–4% and a relative humidity of 30%. The chamber was equipped with recirculating atmosphere filtration system (HePa) to maintain a sterile atmosphere.

Design and Fabrication of SlipChip

SlipChips for chip wash were fabricated as previously reported(2). Photomasks were designed in AutoCAD and ordered from CAD/Art Services, Inc. (Bandon, OR). Soda-lime glass plates with chromium/photoresist and chromium/gold/photoresist coating were purchased from Telic Company (Valencia, CA). The device contained 3,200 microwells for compartmentalization on one plate, and continuous loading channels on the other plate. The depth of wells was 90 µm measured using a profilometer (Dektak 150, Veeco, CA). Through holes were drilled with a 0.035" drill bit (Diamond ball 4F bit, Harvey Tool #74335-C4, Colmar Industrial Supplies, Wheeling, IL) before surface modification. Glass debris from drilling was removed by sonicating the chips in a 1:1 mixture of water and ethanol for more than one hour in a warm water bath. Prior to use, the SlipChips were cleaned with piranha solution, rinsed three times with millipore water followed by 200 proof ethanol, blow dried with nitrogen and silanized with dimethyldichlorosilane using a previously reported protocol (2). An acid piranha solution (caution: this is a corrosive mixture) is used to remove organic contaminants from substrates by mixing 3:1 (v:v) concentrated sulfuric acid (H₂SO₄) with 30% hydrogen peroxide (H₂O₂). Sulfuric Acid (Cat. #A300-212) and 30% Hydrogen peroxide (Cat. #H325-4) can be ordered through Fisher Scientific.

Silanized chips were stored at room temperature in a dessicant box (< 15% humidity, maintained with Drierite). When a glass SlipChip needed to be reused, it was cleaned with Piranha solution first, and then subjected to the same silanization procedure described above.

For the replica-SlipChip, the depth of the features measured by a contact profiler (Dektak 150, Veeco, CA) was 90 µm. The depth was 120 µm for wells, and 60 µm for loading channels of the SlipChip used for depositing PCR reagent wells. The lateral dimensions of cultivation and PCR reagent wells can be measured by photographs from a stereoscope, as shown in the accompanying paper (3).

Performing Chip Wash

The plates of SlipChip were assembled under a layer of tetradecane in a petridish and four binder clips (small binder chips, cat # 429-415, Office Depot) were applied to hold the devices together. The lubricating oil in the loading channel and wells was removed by repeated purging with vacuum for 3 to 5 times at an interval of 1-2 hours between each purging. For anaerobic culture, the SlipChip was placed into the anaerobic chamber for at least 24 hours prior to use. Aqueous solution was loaded onto SlipChip by pipetting, and the device was slipped to form compartments. The solution in the loading channel was removed by purging with a vacuum. In the case of anaerobic cultivation, a gas recirculation pump for atmosphere filtration system was used as a vacuum source. For chip wash experiments with microbial samples, the loading channel can be cleaned by repeated washing with GBSS or PBSc buffer solution to

remove any residual microbial cells and prevent overgrowth of microbes in the loading channel. The continuous loading channels were used as gas exchange channels.

PBS buffer was loaded into the channel to remove the gas phase, and can be used for repeated washing in the case of microbial samples. The SlipChip was designed to collect the chip wash solution with a single outlet (Fig. S1). To collect the solution, 90 μ L PBS buffer was injected into the SlipChip using an Eppendorf pipettor and collected into an Eppendorf pipette tip (1-200 μ L, cat. No. 02-717-141, Fisher Scientific) and transferred into an Eppendorf tube. This process was repeated for three times. 90 μ L tetradeucane was loaded into the SlipChip to further remove the diluted aqueous solution. The solution was also transferred from the pipette tip into the Eppendorf tube.

It is important to keep the lubricating oil in the device from drainage during incubation at elevated temperature or for long-term culture. Loss of lubricating oil in the gap between the two glass plates causes cross-contamination among the confined wells, and cannot be used in the chip wash experiment because the washing buffer would flow through the gap and not into the collection outlet. To prevent loss of oil from the device, a piece of Kimwipe was briefly saturated with 1:1 (vol) mixture of water and tetradeucane and then placed inside a Petri dish. The SlipChip was then placed into the Petri dish and Parafilm was used to seal the Petri dish. The Petri dish was then incubated at the desired temperature for microbial culture.

E. coli Preparation

The green fluorescent protein (GFP)-labeled *E. coli* RP 437 were obtained as a gift from Guillaume Lambert at Princeton University. *E. coli* RP 437 was purchased from CGSC (catalog #: 12122) and transformed with DsRed plasmid.

E. coli cells labeled with DsRed fluorescent proteins were enriched with 50 μ g mL⁻¹ of Ampicillin in LB at 37 °C overnight (12 hours) in a rotary shaking incubator (SI-600 Lab Companion, Jeio Tech) at 200 rpm. Overnight culture was then diluted 100-fold and cultured with 10 μ g mL⁻¹ of Ampicillin and 40 μ mol L⁻¹ IPTG in LB media for 3 hrs. Cells from 1 mL culture were then pelleted at 3000 \times g for 5 min and washed 3 times with 1 mL of ice cold 1× PBS buffer before use.

Quantifying the Recovery Efficiency of Chip Wash

To quantitatively test the recovery efficiency of this method, an aqueous solution of 200 nM Alexa Fluor 488 hydrazide (Invitrogen) in PBS buffer was injected into the device and subsequently collected by chip wash method, normalized to a volume of 500 μ L, and quantified using a fluorospectrometer (Thermo scientific). 0.45 nM, 0.9 nM, 1.8 nM, 3.6 nM and 7.2 nM of Alexa Fluor 488 hydrazide solutions in PBS buffer were used to obtain a calibration curve. The loading volume of the device was calculated to be 18 μ L. Therefore, the concentration of the recovered solution was divided by the concentration of 7.2 nM Alexa Fluor 488 hydrazide to calculate the recovery efficiency.

E. coli cells labeled with DsRed fluorescent proteins were prepared as described above, loaded onto SlipChip and collected immediately using chip wash method. The chip wash solution was normalized to a volume of 500 μ L. 18 μ L of the same solution was diluted to 500 μ L and used as a control. The cells from chip wash solutions as well as the control solution were quantified using INCYTO C-Chip (DHC-N01) under Leica DMI6000 microscope (Leica Microsystems) with a 20 x/0.4NA Leica objective, TX2 filter and a Hamamatsu ORCA-ER camera with 1× coupler with an exposure time of 200 ms. Images were acquired and analyzed by using Metamorph imaging system version 6.3r1 (Universal Imaging). Recovery efficiency was calculated by dividing cell number in the recovered solution by the cell number in the control solution.

Reagents and Equipment for PCR

Primers for PCR were ordered from Integrated DNA Technologies (Coralville, IA). SsoFast EvaGreen Supermix (2X) was purchased from Bio-Rad Laboratories (Hercules, CA). Bovine serum albumin solution (BSA) was purchased from Roche Diagnostics (Indianapolis, IN). PCR Mastercycler and in situ adapter were purchased from Eppendorf (Hamburg, Germany).

Using Chip Wash to Monitor Bacterial Growth with a Single Species Model System

E. coli cells labeled with DsRed fluorescent proteins prepared as described above were serially diluted to a final density of \sim 10⁵ CFU/mL in 10 μ g mL⁻¹ of Ampicillin and 40 μ mol L⁻¹ IPTG in LB media or 1× PBS buffer that does not support growth of bacteria as a negative control and loaded onto SlipChip. SlipChip was incubated at 37 °C overnight. The solution was collected after cultivation using chip wash. Genomic DNA was purified from chip wash solutions using QiaAmp kit (Qiagen). For calibration, genomic DNA was purified from bulk liquid culture of *E. coli* cells labeled with DsRed fluorescent proteins, quantified by Quanti-it DNA high sensitivity quantification kit (Invitrogen), and serially diluted in AE buffer containing 0.01 mg/mL of BSA. The reaction master mixture for qPCR was prepared by mixing 10 μ L of 2X SsoFast EvaGreen Supermix, 1 μ L of forward and reverse primer (10 μ mol L⁻¹), 1 μ L of template solution and 8 μ L of water (Fisher Scientific, BP 2470-1). qPCR was performed on the Eco real-time PCR machine (Illumina, Inc, San Diego, CA) with 27F(4) (5'-AGAGTTGATCCTGGCTCAG -3') and 534R (5'-ATTACCGGGCTGCTGG-3') primers. We observed a 10,000-fold increase in DNA concentration (Fig. S1), suggesting that for this particular model system, non-growing cells contribute to 0.01% of the genetic material recovered from chip wash.

Performing Chip Wash and Plate Wash Experiment with a Two-Species Model System

Cells of *Clostridium scindens* (ATCC 35704) and *Enterococcus faecalis* (ATCC 49532) were separately enriched in Schaedler Anaerobe Broth (Oxoid) at 37°C in an incubator (model# 10-140E, Quincy lab Inc) overnight (~ 16 hours) in anaerobic chamber (Coylab). The culture was diluted 100 fold and incubated at 37°C for 5 hours (*E. faecalis*) and 8 hours (*C. scindens*). The cells were pelleted at 6000 ×g for 3 minutes and washed with PBS for 3 times. Cell numbers of the two species were estimated for loading by cell counting using INCYTO C-Chip (DHC-N01) under Leica DMI6000 microscope (Leica Microsystems) with a 20 x/0.4NA Leica objective. The two species were mixed at 5:1 ratio (*C. scindens*: *E. faecalis*, confirmed by separately plating the two species) in Schaedler Anaerobe Broth at a final density of ~10⁵ CFU/mL, loaded onto SlipChip and incubated at 37°C for 24 hours for growth. Genomic DNA extracted from 18 µL of the same solution containing mixture of cells using QiaAmp kit (Qiagen) was used as non-growth control. To perform plate wash, 4 µL of the same solution containing mixture of cells was plated on Schaedler Anaerobe medium with 2% (wt/vol) noble agar (Fisher Scientific) to achieve a final density of ~300 colonies (counted on day 3 after cultivation when both of the species reached saturation on an agar plate) to reduce interaction between colonies. The cultivar was collected by chip wash and plate wash. Plate wash was performed following a previously described protocol(6) with minor modifications. Cell scrapers (Fisher Scientific) were used to collect cultivar into 1 mL of GBSS buffer. 50 µL of the combined solution was centrifuged at 6000× g for 10 minutes to pellet the cells. Serial dilutions of genomic DNA from macroscale liquid culture of the two species quantified by Quanti-it DNA high sensitivity quantification kit (Invitrogen) were used to calibrate the qPCR machine. The reaction master mixture for qPCR was prepared by mixing 10 µL of 2X SsoFast EvaGreen Supermix, 1 µL of forward and reverse primer (10 µmol L⁻¹), 1 µL of template solution and 8 µL of water (Fisher Scientific, BP 2470-1). For plate wash, ~1 ng/ µL of genomic DNA was used for qPCR as we are interested in the relative ratio of *C. scindens* and *E. faecalis*. qPCR was performed on the Eco real-time PCR machine (Illumina, Inc, San Diego, CA) with ScinF4 (5'-CGTAACGCGCTTTCTTCG-3') and ScinR4 (5'-CCTTCCTCCAGGTTCTCCCT-3') for *C. scindens* and *E. faecalis*_F (5'-CGC TTC TTT CCT CCC GAG T-3') and *E. faecalis*_R (5'-GCC ATG CGG CAT AAA CTG-3'). The two pairs of primers are specific to the targeting species, which was confirmed by qPCR.

We cultivated the two strains separately on SlipChip or on agar plates. To monitor bacterial growth on agar plate, cells were plated on Schaedler Anaerobe Agar separately, incubated at 37 °C and imaged every 24 hours with a Leica MZ 16 stereoscope. The plating experiment was set up with more than three plates for each species. For each time point, one plate was taken out of the anaerobic chamber for imaging and discarded. To image bacterial growth on the SlipChip, bacterial cells from each species were loaded onto a “replica-SlipChip” described in the accompanying paper(3) at a final density of ~10⁵ CFU mL⁻¹ in Schaedler Anaerobe broth. The device was incubated for 24 hours to allow growth of bacteria. The microcolonies were imaged under Leica DMI6000 microscope (Leica Microsystems) with a 20 x/0.7NA Leica objective and a Hamamatsu ORCA-ER camera with 1× coupler under bright field.

On-chip Cultivation of *E. coli* and Splitting of the Microcolonies

E. coli cells were enriched with 50 µg mL⁻¹ of Ampicillin in LB at 34 °C overnight (12 hours) in a rotary shaking incubator at 200 rpm to reach stationary phase. To synchronize cells, overnight culture of each species was then diluted 100-fold and cultured with 10 µg mL⁻¹ of Ampicillin and 40 µmol L⁻¹ IPTG in LB media for 3 hrs. 1 mL culture of cells were then pelleted at 3000 ×g for 5 min and washed 5 times with 1 mL of 1× PBS buffer. Cells were finally suspended in 10 µg mL⁻¹ of Ampicillin and 40 µmol L⁻¹ IPTG in LB media and cell suspension was serially diluted with 10 µg mL⁻¹ of Ampicillin and 40 µmol L⁻¹ IPTG with 0.5% of ultra-low gelling temperature Type IX-A agarose (Sigma-Aldrich) in LB media and mixed to a final density of 2 × 10⁴ and 2 × 10³ CFU mL⁻¹ for *E. coli* strains with GFP and DsRed genes, respectively, and loaded onto replica-SlipChip as described in the accompanying paper(3). SlipChip was incubated at 34 °C for 3 hours and then split into two halves as described in the accompanying paper(3). The bottom half was kept on the holder for colony PCR and the top was preserved at 10 °C on Echo therm chilling plate (Torrey Pines Scientific, Carlsbad, CA) under oil in a Petri dish.

Depositing PCR Reagents on SlipChip

The reaction master mixture was prepared by mixing 100 µL of 2X SsoFast EvaGreen Supermix, 1 µL of forward (DSR_F, 5'-GGACGGCTCCTCATCTACA-3', 100 µmol L⁻¹) and reverse primer (DSR_R, 5'-GGTGATGTCCAGCTTGGACT-3', 100 µmol L⁻¹), 10 µL of 10 µg mL⁻¹ BSA solution, and 68 µL of 2.5% (w/v) ultra-low gelling temperature agarose in water. This mixture was then loaded onto the SlipChip for depositing PCR reagents described in the SI of the accompanying paper (3) by replacing tetradecane in loading channels and this SlipChip for depositing PCR reagent was split to obtain 1,000 droplets deposited on one half of the SlipChip.

Combining the Replica-SlipChip with the PCR Chip

The PCR chip preloaded with PCR reagents was taken off the holder and combined with the bottom piece of the replica chip by aligning through-holes with the pins. A binder clip (5/32' inch capacity, 1/2' inch size, officemax, Itasca, IL) was used to clamp the two plates together, allowing the combined SlipChip to be removed from the holder and the oil without misalignment.

Fluorescence Imaging of PCR Results and *E. coli* with GFP and DsRed Fluorescent Proteins

Fluorescence images were acquired with a Leica DMI6000 microscope (Leica Microsystems) with a 10 x/0.4NA Leica objective and a Hamamatsu ORCA-ER camera with 1× coupler. An L5 filter with an exposure time of 500 ms was used to collect images. For quantitative analysis, fluorescent intensity of a fluorescence reference slide for L5 filter was recorded and used for background

correction. Images were acquired and analyzed by using Metamorph imaging system version 6.3r1 (Universal Imaging) and ImageJ by the National Institutes of Health (<http://rsb.info.nih.gov/ij/download.html>). Processing was applied equally to the entire image.

Preparation of M2GSC Medium

This protocol is adapted from reference (7).

1L M2GSC medium contains the following ingredients:

10 g of casitone, 2.5 g of yeast extract, 4 g of NaHCO₃, 2 g of cellobiose, 2 g of soluble starch, 300 mL of rumen fluid, 2 g of glucose, 1 g of cysteine, 0.45 g of K₂HPO₄, 0.45g of KH₂PO₄, 0.9 g of (NH4)₂SO₄, 0.9 g of NaCl, 0.09 g of MgSO₄ 7H₂O, 0.09 g of CaCl₂, 15 g of Agar Noble and 1 mg of resazurin.

Preparation of M2LC medium

We prepare the basal medium containing: 10 g of casitone, 2.5 g of yeast extract, 4 g of NaHCO₃, 2 g of cellobiose, 2 g of soluble starch, 2 g of glucose, 1 g of cysteine, 0.45 g of K₂HPO₄, 0.45g of KH₂PO₄, 0.9 g of (NH4)₂SO₄, 0.9 g of NaCl, 0.09 g of MgSO₄ 7H₂O, 0.09 g of CaCl₂, 15 g of Agar Noble and 1 mg of resazurin in 700 mL of water.

We mixed 70 µL of the basal medium with 30 µL supernatant of autoclaved luminal sample.

Handling Frozen Stock Solutions of Bacterial Samples in an Anaerobic Chamber

10 µL aliquots of homogenized brush samples and ~50 µL aliquots of homogenized luminal samples were stored in -80 °C freezer. For cultivation, the brush sample was transferred from the freezer to an anaerobic chamber on dry ice with GasPak systems.

Microbial Cultivation from the Biopsy with M2GSC Medium and Performing Plate Wash

An aliquot of cecum brush sample was serially diluted in GBSS buffer. 18 µL of the 10⁴ dilution was plated onto four M2GSC agar plates (4.5 µL for each plate) yielding ~250 colonies per plate after three days of incubation at 37 °C (model# 10-140E, Quincy lab Inc) in an anaerobic chamber (Coylab). Plate wash was performed following a previously described protocol (6) with minor modifications. Cell scrapers were used to collect cultivar into 1 mL of GBSS buffer. The plate wash solutions from four plates were combined into a single tube and mixed by vortexing. 50 µL of the combined solution was centrifuged at 6000× g for 10 minutes to pellet the cells. Genomic DNA was extracted using QiaAmp kit (Qiagen) and quantified by Quanti-it DNA high sensitivity quantification kit (Invitrogen). The experiment was carried out in triplicates. The volume of autoclaved luminal fluid was not enough to prepare plate medium; therefore we did not perform the bulk culture the sample with M2LC medium on agar plates.

Microbial Cultivation from the Biopsy with M2LC and M2GSC Medium and Performing Chip Wash

An aliquot of cecum brush sample was serially diluted in GBSS buffer and then in M2LC. 10⁴ dilution of the brush sample was loaded onto a 3,200 well SlipChip (loading volume of 18 µL and is consistent with sample used the plate wash) and incubated (model# 10-140E, Quincy lab Inc) for three days at 37 °C in an anaerobic chamber (Coylab). Chip wash was performed and the chip wash solution was centrifuged at 6000× g for 10 minutes to pellet the cells. Genomic DNA was extracted using QiaAmp minikit (Qiagen) and quantified by Quanti-it DNA high sensitivity quantification kit (Invitrogen). The experiment was carried out with triplicate devices.

Designing Primers for 16S V1V3 rRNA Gene for Miseq High Throughput Sequencing

We chose high-throughput sequencing for three reasons: First, high-throughput sequencing can be used to profile the community at great depth, and is cost-effective and less labor-intensive than cloning of universally amplified PCR product and Sanger sequencing; Second, compared with testing with primers targeting different groups of microbes, high-throughput sequencing is intrinsically multiplexed and can be used to detect multiple targets simultaneously. This feature is well suited for the “Most Wanted” list, as 45 high priority targets are defined for human gut microbiome. Third, designing and validating primers or probes for specific targets at the resolution of individual OTUs for a complex community from short reads can be challenging for non-experts, and high-throughput sequencing can be used to retrieve the target of interest before the effort for designing target-specific primers.

The primers for variable region V1V3 of 16S rRNA gene was designed similar to that of V4 region of 16s rRNA gene (8, 9). Primer Prospector (10) was used to design the linker region with reference sequences from Greengenes (11) February 2013 release. Possible interactions between barcodes and new pad and linker regions that may yield secondary structures were also screened with Primer Prospector.

16S rRNA Gene Library Construction for Miseq High Throughput Sequencing

The library was prepared according to published protocol. 20 ng of genomic DNA extracted from chip wash solution with M2LC medium or plate wash solutions with M2GSC medium were used in each 50 µL reaction mix. The mixture contained X µL of template DNA (adjusted to 20 ng), (13.5-X) µL of H₂O (Fisher Scientific, BP 2470), 20 µL of 5 Primer Hot MasterMix (5 prime: cat # 2200410), and 1 µL of Primer mix (10µM of Forward primer and barcoded Reverse primer). The PCR reaction was set up in triplicates and PCR product was purified by Agencourt AmPure XP beads (Beckman Coulter Inc, A63881) followed by Qiaquick PCR

purification kit (Qiagen). The purified PCR product was pooled in equal molar quantified by Kapa library quantification kit (Kapa Biosystems, KK4824) and sent for sequencing at GenoSeq Core of UCLA (Los Angeles, CA).

Data Analysis for High Throughput Sequencing of 16S V4 rRNA Gene

OTUs were chosen *de novo* with mothur (12) to identify candidate targets. The results were summarized as an OTU (Operational taxonomic unit) table. The OTU table from chip wash M2LC medium and plate wash with M2GSC medium was sub-sampled to 12599 reads per sample and summarized at genus level using QIIME (13). The relative abundance of each OTU from the two methods is presented in Fig. S4.

Data Analysis for High Throughput Sequencing of 16S V1V3 rRNA Gene

2×250 bp paired end reads were assembled using PANDASeq (14) and de-replicated with usearch (15). The de-replicated reads were sorted by abundance and clustered at 97% identity. Chimeric reads were detected using UCHIME (16) with both *de novo* and reference-based methods. The filtered reads were then searched against the HMP’s “Most Wanted” list for targets within the identity of 97% using usearch v6.0.293. Alignment of the sequence from chip wash with the sequence of OTU158 from the most wanted list is shown in Fig. S5. Detailed scripts are provided in the SI below.

Using qPCR to Quantify Bacterial Genomic DNA from the Total Bacteria and OTU158

Primers for PCR were ordered from Integrated DNA Technologies (Coralville, IA). SsoFast EvaGreen Supermix (2X) was purchased from Bio-Rad Laboratories (Hercules, CA). 1.5 ng purified gDNA was used to prepare the reaction mixture of a total volume of 30 μ L. Water (Fisher Scientific, BP 2470) was used as negative control. Universal primers for the V4 region of 16S rRNA gene (515F, 5'-GTGCCAGCMGCCGCGTAA-3', 806R, 5'-GGACTACHVGGGTWTCTAAT-3') were used to quantify total bacterial load, and OTU158 specific primers (158F, 5'-AGA ATC TAC TGA AAG AGT TTT CGG A-3', 158R, 5'-TTC TAG AGG TAC CGT CTT CTG CT-3') were used to quantify the concentration of OTU158. The mixture was split into 3 aliquots and loaded onto the Eco real-time PCR machine (Illumina, Inc, San Diego, CA). Reactions were incubated for 2 min at 98 °C, followed by 40 cycles of 5 s at 98 °C, 3 s at 60 °C. Data analysis was performed using Eco software.

Using Splitting Technology and the On-chip PCR Method to isolate the bacterium: isolate microfluidicus 1

An aliquot of cecum brush sample was serially diluted in GBSS buffer and then in M2LC medium. 10^4 dilution of the brush sample was loaded onto a 1,000 well replica-SlipChip (3) and incubated (model# 10-140E, Quincy lab Inc) for two days at 37 °C in an anaerobic chamber (Coylab). The replica-SlipChip was split as described in the accompanying paper (3), and on-chip PCR was performed and analyzed as described above with the following modification: The PCR reaction master mixture was prepared by mixing 150 μ L of KAPA 2G Robust Hot Start Readymix (KAPA BIOSYSTEMS), 1.5 μ L of forward and 1.5 μ L of reverse primer (100 μ mol L⁻¹), 15 μ L of 10 μ g mL⁻¹ BSA solution, and 94.5 μ L of 2.5% (w/v) ultra-low gelling temperature agarose in water, and 7.5 μ L of 40X SYBR green (Sigma-Aldrich). The staining of cell material with SYBR green was observed and could be used to estimate the number of microcolonies grown on device could be estimated. Microbes from the PCR-positive wells on the plate for sample preservation was retrieved as described in the accompanying paper (3), and plated on M2GSC plate. After two days of incubation, the cluster of colonies was used for both colony PCR and streaking additional plates.

16S rRNA Gene Analysis for isolate microfluidicus 1

Genomic DNA for PCR amplification was isolated using QiaAmp kit (Qiagen) following the manufacturer’s protocol with the following modification. We added a bead-beating step using lysing matrix B (MP Biomedicals 6911-500) that was shaken using a Mini-Beadbeater-16 (BioSpec Products, Inc.) for 1 min. The 16S rRNA gene was amplified by PCR using AccuPrimer Pfx DNA polymerase (Invitrogen). Primers 8F (5'-AGAGTTGATCCTGGCTCAG-3') and 1492R (5'-GGTTACCTGTTACGACTT-3') (17) were used for PCR amplification. PCR amplification was performed by Biorad thermocycler with 2 min incubation at 95 °C, followed by 34 cycles at 95 °C for 15s, 55 °C for 30s and 68 °C for 90s. Amplified PCR product was cloned into TOPO vector (Invitrogen) and transformed into TOPO10 *E. coli* cells (Invitrogen) on LB/Amp+ medium. The plates were incubated at 37 °C overnight and single colonies were picked for liquid culture. Plasmids were purified from cells using Qia miniprep kit (Qiagen). Plasmid DNA was then amplified by PCR with the same protocol as described above using M13F/M13R primers (Invitrogen). PCR products were purified using Qia quick PCR purification kit (Qiagen).

Sequencing PCR Products and Data Analysis

PCR products were sequenced by Laragen, Inc. (Culver City, CA) using T3 and T7 as sequencing primers (Invitrogen). The paired-end reads were assembled in Seqman Pro (DNASTAR) and manually trimmed to remove the adapters and PCR primers in Microsoft Word. 15 assembled sequences were aligned using muscle (18) and usearch (15).

TEM

TEM was performed in the Jensen laboratory electron microscopy facility at the California Institute of Technology with 200 mesh formvar/ carbon grids on TECNAI 120 keV TEM (FEI, Hillsboro, OR) equipped with a Gatan 2k by 2k CCD camera for image acquisition.

Optical Microscopy

Optical microscopy of the isolate was obtained by suspending the cells in PBS buffer and imaged using a 63×/1.2 NA Leica objective with a Leica DMI6000 microscope (Leica Microsystems) and a Hamamatsu ORCAER camera.

Ribosomal Database Project (RDP) classification of 16S rRNA gene of OTU158

Taxonomic assignment of the sequencing results for OTU158 (isolate microfluidicus 1) was generated using the online RDP classifier with 16S rRNA training set 9. The results are summarized below:

Classification on sequence of OTU158 retrieved from high throughput sequencing of V4 region of 16S rRNA gene:
Bacteria(100);Firmicutes(100);Clostridia(100);Clostridiales(100);Ruminococcaceae(100); Oscillibacter(100);

Classification on sequence of OTU158 retrieved from high throughput sequencing of V1V3 region of 16S rRNA gene:
Root(100);Bacteria(100);Firmicutes(100);Clostridia(100);Clostridiales(100);Ruminococcaceae(100);Oscillibacter(99);

Classification on sequence of two types of full length 16S rRNA gene of isolate microfluidicus 1 from Sanger sequencing:
Root(100%) Bacteria(100%) Firmicutes(100%) Clostridia(100%) Clostridiales(100%) Ruminococcaceae(100%) Oscillibacter(75%);
Root(100%) Bacteria(100%) Firmicutes(100%) Clostridia(100%) Clostridiales(100%) Ruminococcaceae(100%) Oscillibacter(84%);

Numbers in parentheses give the classification confidence level; threshold is 80%.

Phylogeny

Phylogenetic analyses of 16S rRNA gene sequences were performed using the software package ARB (19) as well as MrBayes (20). Automatic alignments of sequences obtained from our culture as well as from reference strains were manually refined within ARB. A consensus tree was constructed based on maximum-likelihood (ML) calculation (using the Hasegawa, Kishino and Yano substitution model), and by collapsing all nodes with parsimony bootstrap (5,000 iterations) support ≤50% or Bayesian support below ≤70%. Conditions for Bayesian inference were as follows: 2 parallel runs; 1,000,000 tree generations; sample frequency 100; final split frequency 0.007; potential scale reduction factor 1.006; burnin of 25% of sample. For phylogenetic analyses only sequences of cultured members of the *Clostridia* for which the 16S rRNA sequence was available were considered. Using a manually designed sequence filter we excluded highly variable in-del positions from the analysis, resulting in 1,371 alignment positions for tree calculations.

Fluorescence *in situ* Hybridization

16S rRNA targeted FISH was carried out following established protocols (21). In brief, formaldehyde- and ethanol-fixed samples were hybridized at 46°C with FAM- and Cy3-labeled oligonucleotide probes for 16 hours in a formamide-containing humid chamber. To test whether cell wall digestion leads to an increase in fluorescence detection and/or labeling intensity, before hybridization samples were pre-treated with either (i) 10 mg mL⁻¹ lysozyme in TE buffer (1 h at 37°C in a humid chamber); (ii) 15 µg mL⁻¹ proteinase K in TE buffer (10 min at room temperature, *i.e.* 23°C) followed by 0.01 M HCl (10 min at 23°C); or (iii) a 1:1 mix of acetone:methanol (15 min at 23°C). Formamide concentrations in the hybridization buffer were as recommended: 20–35% for probe mix EUB338 I-III (22, 23) and control probe NonEUB338 (24); 35% for probe Arch915 (25); 20% for probe EUK516 (22). The two newly designed probes Clostr183-I (AAAGAT CAT GCG ACCTCT) and Clostr183-II (AAGAAT CAT GCG ACCCCT) were hybridized at 15% (at concentrations >20% we did not observe any fluorescence signal). Via competition for the same binding site, these probes are able to distinguish between the two 16S rRNA gene sequence types obtained from our culture. After hybridization, slides were washed for 10 min in pre-warmed washing buffer at 48°C. Then, they were dipped into pre-cooled deionized water (4°C) and dried using pressurized air. Slides were mounted with DAPI/Citifluor and analyzed using an Olympus BX51 epifluorescence microscope. Fluorescence images were analyzed using the software provided by the microscope manufacturer and ImageJ by the National Institutes of Health (<http://rsb.info.nih.gov/ij/download.html>). No unspecific labeling was observed when control probe NonEUB338 (9) was applied to our samples.

Whole Genome Sequencing

The genome of isolate microfluidicus 1 was sequenced on Illumina Hiseq 2000 and assembled using a combination of velvet (26) and GapCloser (27). We obtained 83 contigs with N50 length of 131 kbp.

Scripts for Identifying Targets from “Most Wanted” list from paired-end Miseq reads

```
File name: get_target_all
#!/bin/bash
#file created on Dec 2, 2012
#Author= "Liang Ma"
#Email= liangma.chem@gmail.com
# usage example: get_target_all ~/Downloads/reads/
#set environmental variables
#usearch program
export u=/Users/Liang/scripts/usearch/usearch6.0.293_i86osx32_lm
#reference database for UCHIME
export UCHIME_REFUDB=/Users/Liang/scripts/Database/gold.fa
# fasta file of consensus sequence from most wanted list, high priority group
export MostWanted=/Users/Liang/scripts/Database/MostWanted.fa
export scriptsHome=/Users/Liang/scripts
# $1 is the folder containing fastq files with paired end reads
cd $1
for file in $(ls| grep .*R1.*\.fastq)
do
FN=$(echo $file|grep -o .*R)
FileR="$FN"2_001.fastq
#assemble reads with pandaseq
pandaseq -f $file -r $FileR -o 10 > "$FN.fa"
$u -derep_fulllength $FN.fa -output "derep_$FN.fa".fa --sizeout
readname='results'
mkdir -p $FN$readname
cd $FN$readname
$scriptsHome/get_target1 ../"derep_$FN.fa.fa"
cd $1
done
```

```
File name: get_target1
#!/bin/bash
#file created on Dec 2, 2012
#Author= "Liang Ma"
#Email= liangma.chem@gmail.com
# usage example: get_target1 reads.fa
if [ x$1 == x ] ; then
    echo Missing FASTA filename >> /dev/stderr
    exit 1
fi
if [ x$UCHIME_REFUDB == x ] ; then
    echo Must set \$UCHIME_REFUDB >> /dev/stderr
    exit 1
fi
if [ x$MostWanted == x ] ; then
    echo Must set \$MostWanted >> /dev/stderr
    exit 1
fi
# Sort by decreasing size
$u -sortbysize $1 -output bysize.fa
#cluster at 97% identity
$u -cluster_smallmem bysize.fa -id 0.97 -sizein -sizeout -centroids 97.fa
# Chimera filter with UCHIME de novo
$u -uchime_denovo 97.fa -chimeras ch.fa -nonchimeras nonch.fa -uchimeout denovo.uchime
# Chimera filter with UCHIME ref
$u -uchime_ref nonch.fa -db $UCHIME_REFUDB -nonchimeras filtered.fa -strand plus -uchimeout ref.uchime
# find hits with MostWanted list
$u -usearch_global filtered.fa -db $MostWanted -id 0.97 -strand plus -fastapairs mostwanted.fastapairs
```

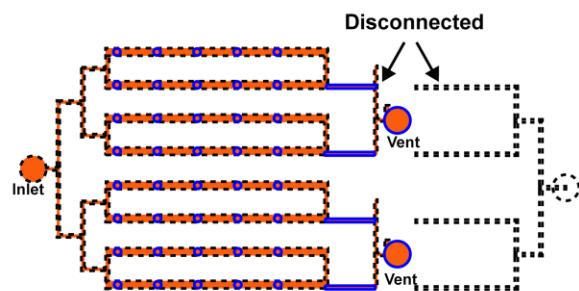
> Sequence of rrnA of 16S rRNA gene of isolate microfluidicus 1

GACGAACGCTGGCGCGTCTAACACATGCAAGTCGAACGAGAACTACTGAAGGAGGATTGCTCAACGGAAGTAGAGGAAAGTGGCGGACGGGT
GAGTAACGCGTGAGGAACCTGCCTGAAGAGGGGACAACAGTTGAAACAGACTGCTAATACCGCATGACGCATAGGGGTCGCATGATTCTATGCCA
AAGATTITATCGCTCAAGATGGCTCGCGTCTGATTAGCTAGTTGGGGGTAACGGCCCACCAAGGGCAGCAGTACGTAAGCCGGACTGAGAGGTTGAA
CGGCCACATTGGGACTGAGATAACGGCCCAGACTCCTACGGGAGGCAGTGGGAATATTGGCAATGGGCGAAGCCTGACCCAGCAACGCCGCG
TGAAGGAAGAAGGCTTCGGTTGAAACTCTTAAAGAGGGAAGAGCAGAAGACGGTACCTCTAGAATAAGCCACGGCTAACTACGTGCCAGCAGC
CGCGGTAACTACGTAGGTGGCAAGCGTTGCGGATTACTGGGTGAAAGGGCTGAGCCGGAGGTTGACATGGGCTGCAAGTCAGATGTGAAATCCATGGGCTCAAC
CCATGAACGCTCATTTGAAACTGTAGATCTTAGTGTGGGAGGCAATCGGAATTCTAGTGTAGCGTAAATGCGTAGATATTAGGAGGAACACCA
GTGGCGAAGGCGGATTGCTGGACGATAACTGACGGTGGGAGGCAAGCTGAGGATTAGATACCCCTGGTAGTCCACACTGTAAC
ATGAATACTAGGTGTGCGGGACTGACCCCTGCGTGGGAGGCAACTGAGGAGGAAAGCAGAAGCCGAGGAGGTTGACATGGGCTGCAAGGTTGAAACTCAAAG
GAATTGACGGGGGCCCCACAAGCGGTGGATTATGTGGTTAATTCAAGCAACGCAACGGGAGGAAAGCAGGAGGAGGTTGACATCCCTGCTAACGAGC
GATACATTAAGGTGCCCTCGGGAAAGCAGAGACAGGTGGTGCATGGGTGCTGAGCTGGTAGTGGGAGGAGGTTGACATCCCTGCTAACGAGC
AACCCCTATTGTTAGTTGCTACGCAAGAGCACTCTAGCGAGACTGCCGTGACAAAACGGAGGAAAGGTGGGAGCAGCTCAAATCATGCCCTTA
TATCCTGGGCTACACCGTAATAACATGGCGTCAACAGAGGGAAGCAGGAGGAGGAGGAAAGCAGGAGGAGGTTGACATCCCTGCTAACGAGC
AGGCTGCAACTCGCCTGATGAAGTCGAATCGCTAGTAATCGCGGATCAGCATGCCGCGTGAATACGTTCCGGCCTTGTACACACCGCCCGTCA
CACCATGAGAGTCGGAACACCGAAGTCGTAGCCTAACCTGAAAAGGAGGGCGCGGCCGAAGGTGGGTCGATAATTGGGTTG

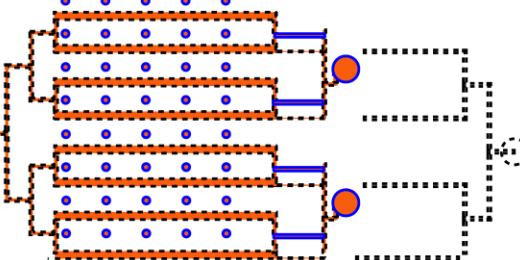
> Sequence of rrnB of 16S rRNA gene of isolate microfluidicus 1

GACGAACGCTGGCGCGTCTAACACATGCAAGTCGAACGAGAACTACTGAAAAGAGTTTCCGACAATGGAAGTAGAGGAAAGTGGCGGACGGGT
GAGTAACGCGTGAGGAACCTGCCTGAAGAGGGGACAACAGTTGAAACAGACTGCTAATACCGCATGACGCATAGGGTCGCATGATCTTATGCCA
AAGATTITATCGCTCAAGATGGCTCGCGTCTGATTAGCTAGTTGGGGGTAACGGCCCACCAAGGGCAGCAGTACGTAAGCCGGACTGAGAGGTTGAA
CGGCCACATTGGGACTGAGATAACGGCCCAGACTCCTACGGGAGGCAGTGGGAATATTGGCAATGGGCGAAGCCTGACCCAGCAACGCCGCG
TGAAGGAAGAAGGCTTCGGTTGAAACTCTTAAAGAGGGAAGAGCAGAAGACGGTACCTCTAGAATAAGCCACGGCTAACTACGTGCCAGCAGC
CGCGGTAACTACGTAGGTGGCAAGCGTTGCGGATTACTGGGTGAAAGGGCGTGCAGCCGGGCTGCAAGTCAGATGTGAAATCCATGGGCTCAAC
CCATGAACGCTCATTTGAAACTGTAGATCTTAGTGTGGTGCAGGCGGGCAATCGGAATTCTAGTGTAGCGTAAATGCGTAGATATTAGGAGGAACACCA
GTGGCGAAGGCGGATTGCTGGACGATAACTGACGGTGAGGCGCAGAAGTGTGGGAGGAAACAGGATTAGATACCCCTGGTAGTCCACACTGTAAC
ATGAATACTAGGTGTGCGGGACTGACCCCTGCGTGGGAGGCAACTAAACACAATAAGTATTCCACCTGGGAGTACGATCGCAAGGTTGAAACTCAAAG
GAATTGACGGGGGCCCCACAAGCGGTGGATTATGTGGTTAATTCAAGCAACGCAACGCAAGAACCTTACCAAGGGTTGACATCCCTGCTAACGAGT
GATACATTAAGGTGCCCTCGGGAAAGCAGAGACAGGTGGTGCATGGGTGCTGAGCTGGTAGTGGGAGGTTAAGTCCCGCAACGAGC
AACCCCTATTGTTAGTTGCTACGCAAGAGCACTCTAGCGAGACTGCCGTGACAAAACGGAGGAAAGGTGGGAGCAGCTCAAATCATGCCCTTA
TATCCTGGGCTACACCGTAATAACATGGCGTCAACAGAGGGAAGCAGGAGGAGGAGGAAAGCAGGAGGAGGTTGACATCCCTGCTAACGAGC
AGGCTGCAACTCGCCTGATGAAGTCGAATCGCTAGTAATCGCGGATCAGCATGCCGCGTGAATACGTTCCGGCCTTGTACACACCGCCCGTCA
CACCATGAGAGTCGGAACACCGAAGTCGTAGCCTAACCTGAAAAGGAGGGCGCGGCCGAAGGTGGGTCGATAATTGGGTTG

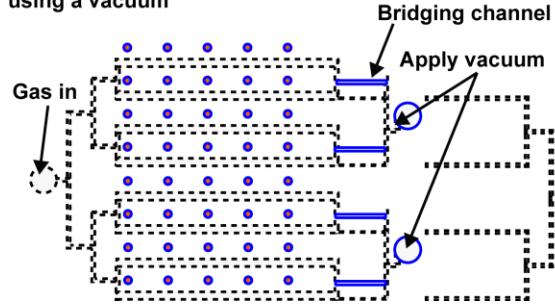
a) Loading of aqueous solution



b) Slipping the blue/solid layer upward for compartmentalization



c) Removing residual aqueous solution using a vacuum



d) Slipping the blue/solid layer downward and then to the right to enable chip wash

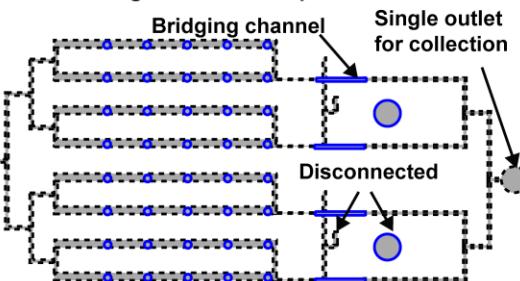


Fig. S1 Illustration of operation of the device showing how the bridging channels, vents and outlet for collection were used during chip wash. This schematic drawing uses a simplified design with fewer wells to illustrate the principle.

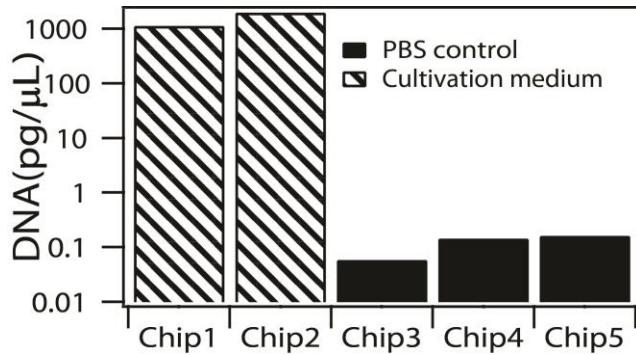


Fig. S2 Validation of the chip wash method with a model microorganism, *E. coli*. *E. coli* cells were loaded onto SlipChip with either medium for cultivation or a PBS buffer that does not support growth of cells. The cultivar was collected by chip wash method and DNA was extracted and quantified using qPCR. A 10,000-fold increase in the amount of DNA was observed between the condition that supports growth of *E. coli* vs. the non-growth control.

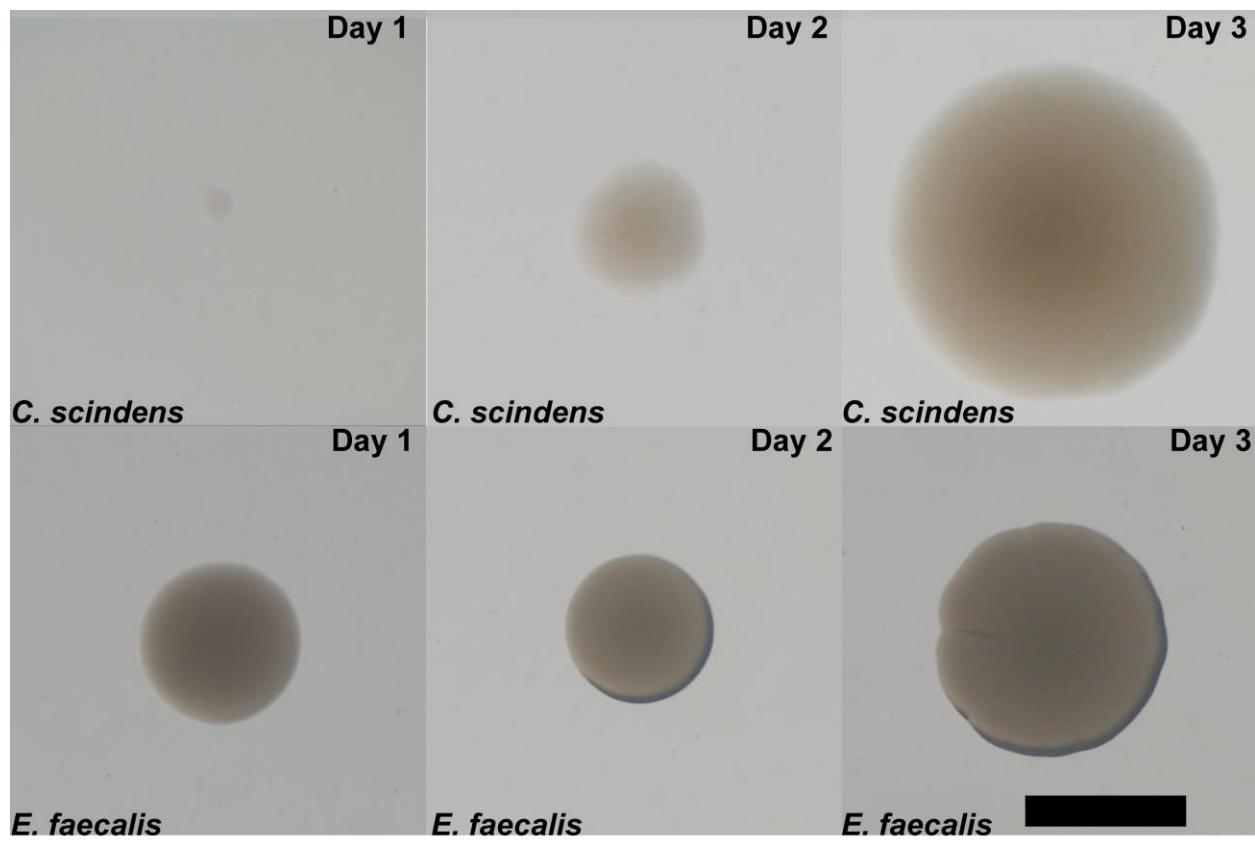
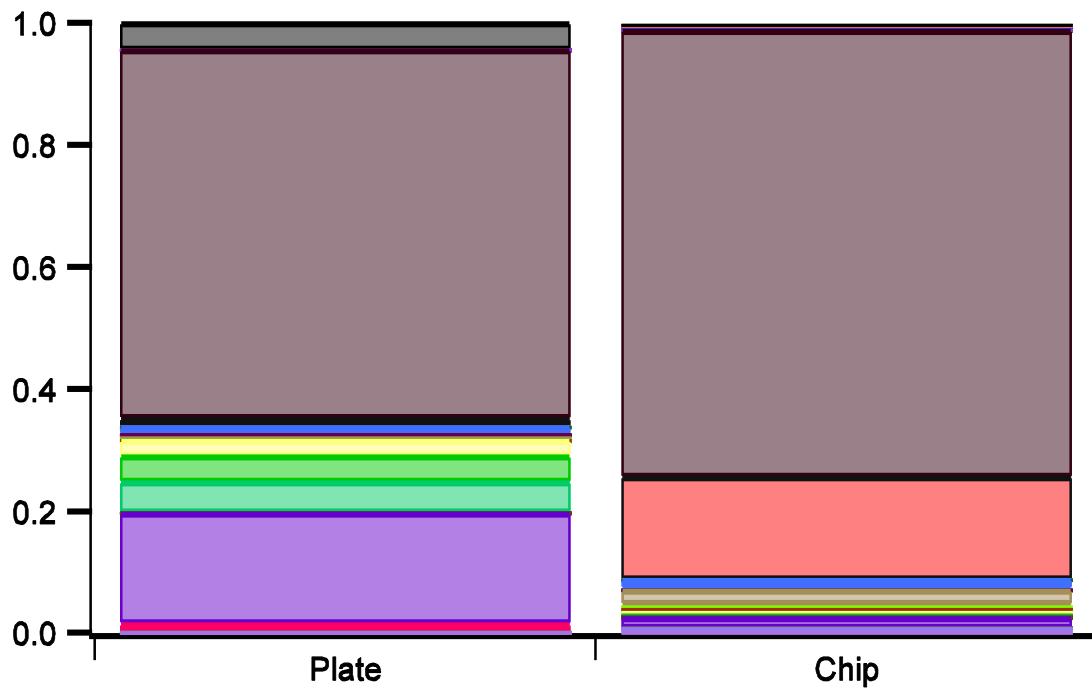


Fig. S3 Photograph showing time series of growth of *Clostridium scindens* and *Enterococcus faecalis* on agar plate. Scale bar is 1 mm.



- Bacteria;Actinobacteria;Actinobacteria;Bifidobacteriales;Bifidobacteriaceae;Bifidobacterium
- Bacteria;Actinobacteria;Actinobacteria;Coriobacteriales;Coriobacteriaceae;Collinsella
- Bacteria;Actinobacteria;Actinobacteria;Coriobacteriales;Coriobacteriaceae;Eggerthella
- Bacteria;Actinobacteria;Actinobacteria;Coriobacteriales;Coriobacteriaceae;Gordonibacter
- Bacteria;Bacteroidetes;Bacteroidia;Bacteroidales;Bacteroidaceae;Anaerorhabdus
- Bacteria;Bacteroidetes;Bacteroidia;Bacteroidales;Bacteroidaceae;Bacteroides
- Bacteria;Bacteroidetes;Bacteroidia;Bacteroidales;Porphyromonadaceae;Parabacteroides
- Bacteria;Bacteroidetes;Bacteroidia;Bacteroidales;unclassified;unclassified
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;Anaerostipes
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;Blautia
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;Clostridium_XIVa
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;Clostridium_XIVb
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;Coprococcus
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;Dorea
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;Lachnospiracea_incertae_sedis
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;Roseburia
- Bacteria;Firmicutes;Clostridia;Clostridiales;Lachnospiraceae;unclassified
- Bacteria;Firmicutes;Clostridia;Clostridiales;Ruminococcaceae;Faecalibacterium
- Bacteria;Firmicutes;Clostridia;Clostridiales;Ruminococcaceae;Flavonifractor
- Bacteria;Firmicutes;Clostridia;Clostridiales;Ruminococcaceae;Oscillibacter
- Bacteria;Firmicutes;Clostridia;Clostridiales;Ruminococcaceae;Subdoligranulum
- Bacteria;Firmicutes;Clostridia;Clostridiales;Ruminococcaceae;unclassified
- Bacteria;Firmicutes;Clostridia;Clostridiales;unclassified;unclassified
- Bacteria;Firmicutes;Erysipelotrichia;Erysipelotrichales;Erysipelotrichaceae;Clostridium_XVIII
- Bacteria;Firmicutes;Erysipelotrichia;Erysipelotrichales;Erysipelotrichaceae;Coprobacillus
- Bacteria;Firmicutes;Erysipelotrichia;Erysipelotrichales;Erysipelotrichaceae;unclassified
- Bacteria;Firmicutes;Negativicutes;Selenomonadales;Veillonellaceae;Dialister
- Bacteria;Firmicutes;unclassified;unclassified;unclassified;unclassified
- Bacteria;Proteobacteria;Gammaproteobacteria;Pseudomonadales;Pseudomonadaceae;Pseudomonas
- Bacteria;Proteobacteria;Gammaproteobacteria;Xanthomonadales;Xanthomonadaceae;Silanimonas
- Bacteria;unclassified;unclassified;unclassified;unclassified

Fig. S4 Bar chart of relative abundance of OTUs grouped at genus level from 16S rDNA V4 high throughput sequencing of chip wash cultivar with M2LC and plate wash cultivar with M2GSC. We observed that more *Clostridium XIII* and *Bifidobacterium* can be observed from the plate wash solution, while some members such as *Gordonibacter*, *Anaerostipes*, *Oscillibacter* and *Silanimonas* can only be observed from chip wash method.

ACGCTGGCGCGTGTAAACACATGCAAGTCGAACGAGAACTACTGAAAGAGTTTCGG
ACGCTGGCGCGTGTAAACACATGCAAGTCGAACGAGAACTACTGAAAGAGTTTCGG
ACAATGGAAGTAGAGGAAAGTGGCGACGGGTGAGTAACGCGTGAGGAACCTGCCTGAA
ACAATGGAAGTAGAGGAAAGTGGCGACGGGTGAGTAACGCGTGAGGAACCTGCCTGAA
GAGGGGGACAACAGTTGAAACGACTGCTAATACCGCATGACGCATAGGGTTCGATGAT
GAGGGGGACAACAGTTGAAACGACTGCTAATACCGCATGACGCATAGGGTTCGATGAT
TTTATGCCAAAGATTATCGCTAAAGATGGCTCGCGTCTGATTAGCTAGTTGGTGGG
CTTATGCCAAAGATTATCGCTCAAGATGGCTCGCGTCTGATTAGCTAGTTGGCGGG
GTAACGGCCCACCAAGGCAGCATCAGTAGCCGACTGAGAGGTTGAACGGCCACATTGG
GTAACGGCCCACCAAGGCAGCATCAGTAGCCGACTGAGAGGTTGAACGGCCACATTGG
GACTGAGATA CGGCCAGACT CCTACGGGAGGCAGCAGTGGGAATTGGGCAATGGG
GACTGAGATA CGGCCAGACT CCTACGGGAGGCAGCAGTGGGAATTGGGCAATGGG
GCAAGCCTGACCCAGAACGCCGCGTGAAGGAAGAAGGTTGGGTTGAAACTTCTT
GCAAGCCTGACCCAGAACGCCGCGTGAAGGAAGAAGGTTGGGTTGAAACTTCTT
TAAGAGGGAAGAGCAGAACAGCGTACCTCTAGAATAAGCCACGGCTAACACTCGT
TAAGAGGGAAGAGCAGAACAGCGTACCTCTAGAATAAGCCACGGCTAACACTCGT

TOP: Consensus sequence from member OTU_158_V1V3 of Most Wanted list
BOTTOM: Consensus sequence from chip wash 16s rDNA V1V3 survey

Fig. S5 Identifying OTU158 from high throughput sequencing of V1V3 region of 16S rRNA gene of the chip wash solution. The sequence retrieved from high throughput sequencing aligned well with the consensus sequence of OTU158 from HMP's "Most Wanted" list.

GGGT GAGT AACCGT GAGGAACCT GCCT TGAAGAGGGGACAACAGTT GAAAACGACT GC
GGGT GAGT - ACGCGT GAGGAACCT GCCT TGAAGAGGGGACAACAGTT GAAAACGACT GC
TAAT ACCGCAT GACGCAT AGGGGT CGCAT ATT TT ATGCCAAAGATT TAT CGCT GAAAG
TAAT ACCGCAT GACGCAT AGAGGT CGCAT GAT CTT ATGCCAAAGATT TAT CGCTT CAAG
ATGGCCT CGCGT CT GATT AGCT AGT GTT GGGT AACGGCCACCAAGGC GACGAT CAGT
ATGGCCT CGCGT CT GATT AGCT AGT GTT GGGGGT AACGGCCACCAAGGC GACGAT CAGT
AGCCGGACT GAGAGGT GAACGGCCACATTGGGACT GAGAT ACGGCCCAGACT CCTACGG
AGCCGGACT GAGAGGT GAACGGCCACATTGGGACT GAGAT ACGGCCCAGACT CCTACGG
GAGGCAGCACT GGGGAAT ATTGGCAATGGGCAGCAAGCT GACCCAGCAACGCCGCTGA
GAGGCAGCACT GGGGAAT ATTGGCAATGGGCAGCAAGCT GACCCAGCAACGCCGCTGA
AGGAAGAAGGCTT CGGGTT TAAACTTCTTTAAAGA
AGGAAGAAGGCTT CGGGTT TAAACTTCTTTAAAGA

TOP: Consensus sequence from member OTU_158_V1V3 of Most Wanted list
BOTTOM: Partial sequence from chip wash PCR with primers targeting OTU 158 V1V3

Fig. S6 Validating species specific primers for OTU158 by Sanger sequencing. The sequence retrieved from Sanger sequencing aligned well with the consensus sequence of OTU158 from HMP’s “Most Wanted” list.

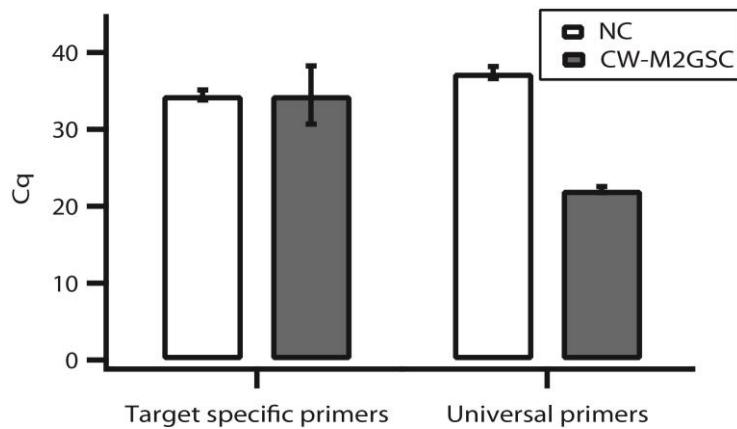


Fig. S7 qPCR with target specific primers (left) showing that the target was not present in chip wash solution of M2GSC nor in the blank negative control; Universal primers of 16s rDNA (right) showed that both chip wash contained bacterial genomic DNA and had a lower Cq value than the blank negative control. NC is negative control, and CW-M2GSC is chip wash with M2GSC medium.

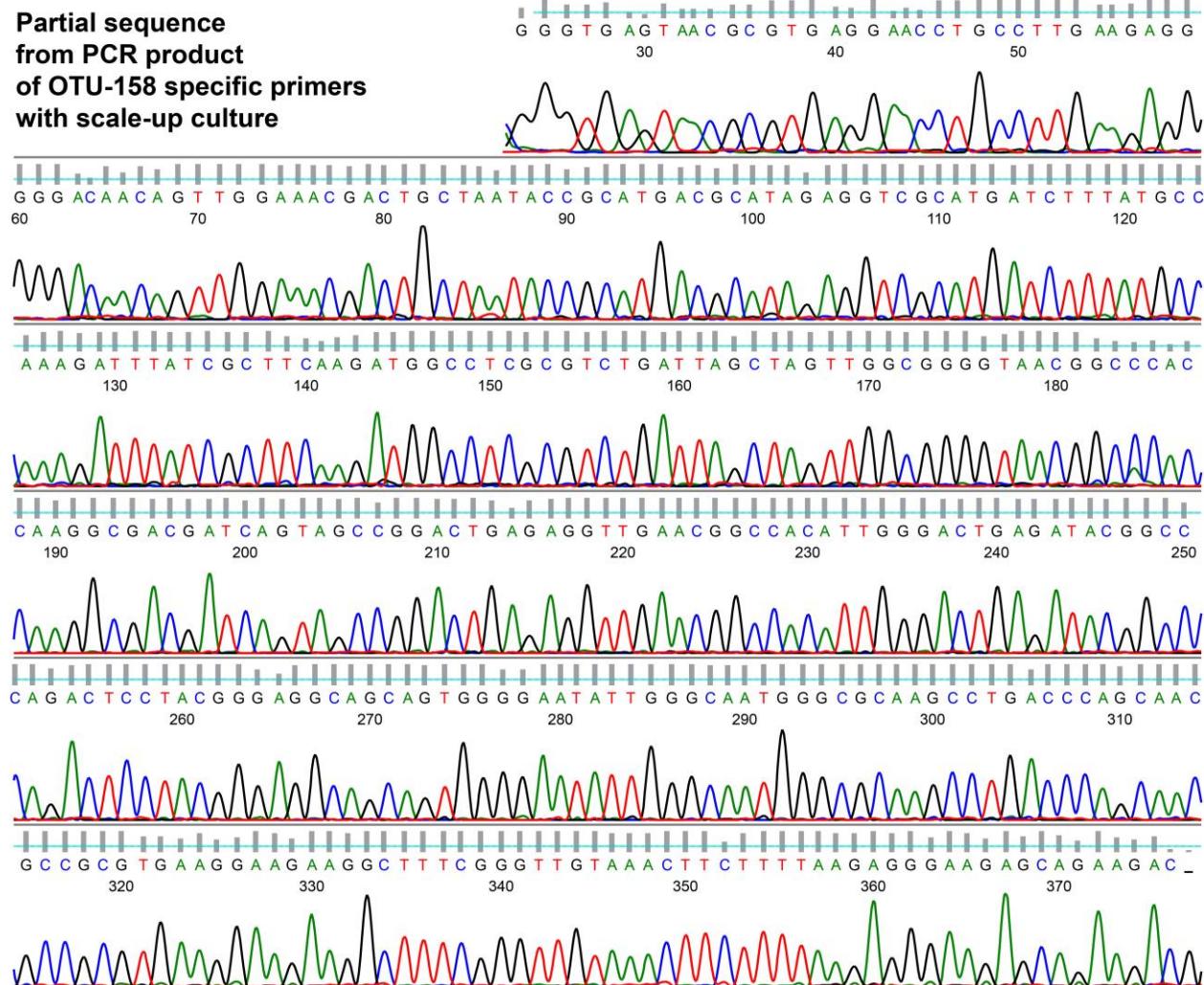


Fig. S8 Chromatogram from Sanger sequencing of positive PCR product of the first scale-up culture with target-specific primers.

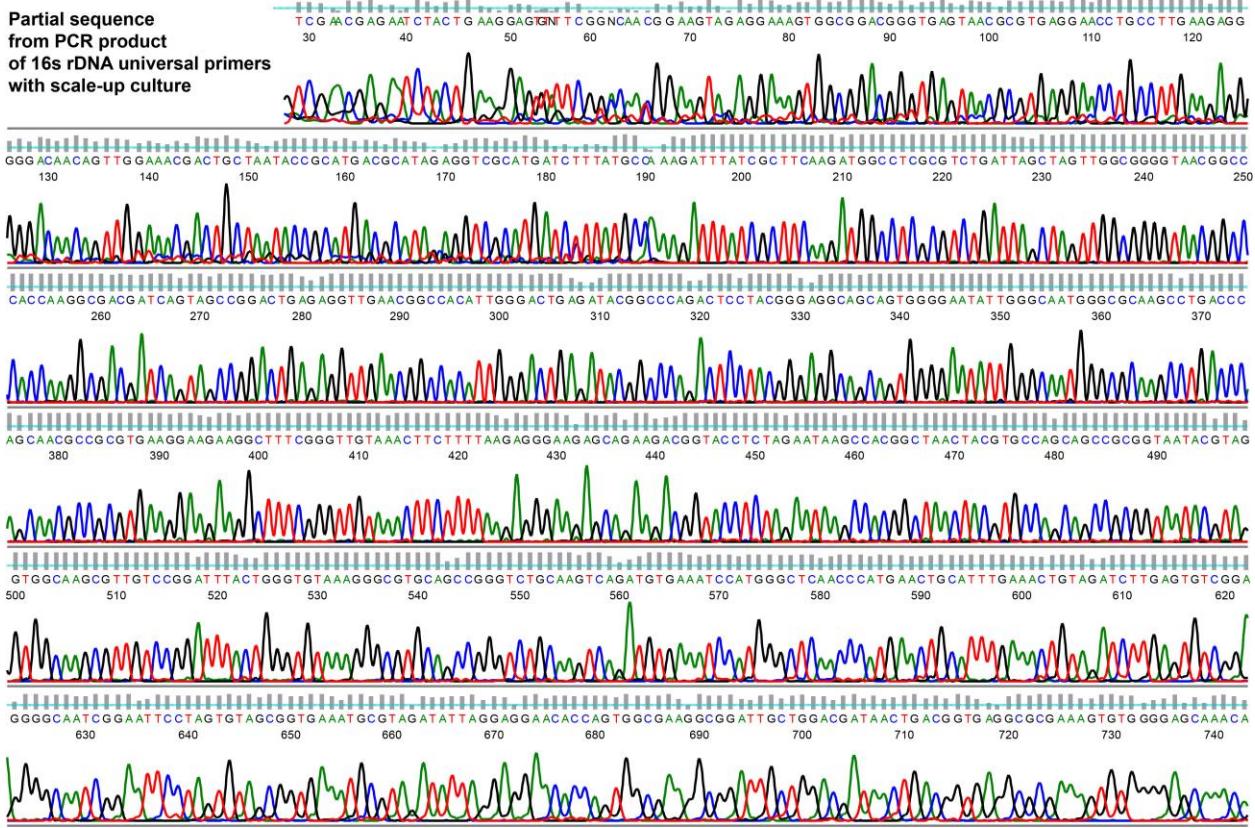


Fig. S9 Validating the scale-up colonies from the splitting SlipChip approach by Sanger sequencing of the positive PCR product of the first scale-up culture using universal primers. No contamination was observed, with minor heterogeneity at the beginning of the chromatogram.

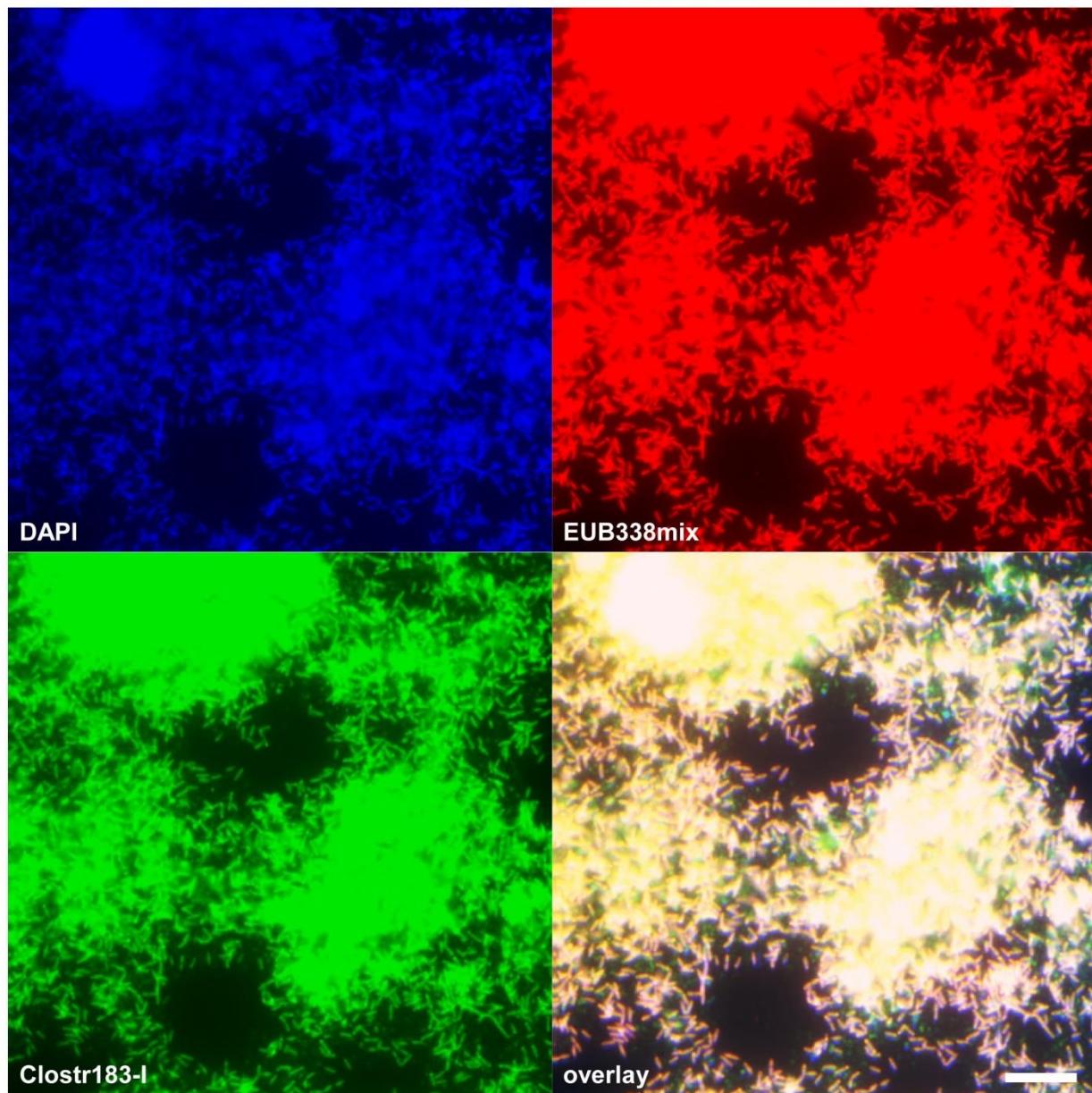


Fig. S10 FISH visualization of bacterial cells within pure culture of isolate microfluidicus 1. All cells detected by the general bacterial probe mix EUB338 I-III are also detected via the newly designed Clostr183-I probe, targeting isolate microfluidicus 1. While some cells (as identified by DAPI) did not bind any of these two probes, no archaea or eukaryotes were detected. Thus, FISH-negative cells are likely sporulating cells with substantially decreased ribosome content as compared to log-phase cells. Scale bar is 10 μ m.

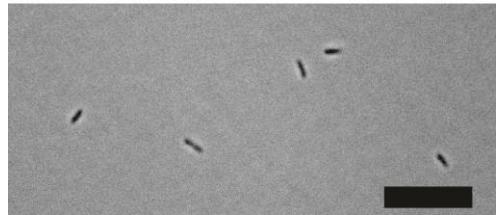


Fig. S11 Optical microscopy of isolate microfluidicus 1. Scale bar is 20 μm .

Detailed Contribution Description

LM and RFI designed the experiment as described in Figure 1. LM designed, performed experiments, and analyzed the data in Figures 2, 4, and 5. LM and JK designed, performed experiments, and analyzed the data in Figure 3. RH designed and performed FISH experiments and phylogenetic analyses for Figure 6, wrote the corresponding Results and Discussion and Supporting Information sections, and helped to edit the paper. NH was involved in clinical specimen acquisition and preparation in Figure 5A. IH performed all procedures for clinical specimen acquisition in Figure 5A. EC designed the study and clinical operations in Figure 5A. MK helped with device infrastructure and performed the experiment in SI Figure 11.

Table S1

#metadata of OTU_158_V1V3 and OTU_896_V1V3, retrieved from http://www.hmpdacc.org/most_wanted/ on Apr 27, 2012

otuID	otu_158_V1V3	otu_896_V1V3
priorityGroup	HIGH PRIORITY/MOST WANTED	HIGH PRIORITY/MOST WANTED
variableRegion	V1V3	V1V3
count454	14888	788
consensusLength	475	463
	ACGCTGGCGGCGTCTAACACATGCAAGTCGA ACGAGAACTACTGAAAGAGTTTCGGACAATG GAAGTAGGGAAAGTGGCGGACGGGTGAGTA ACGGTGAAGAACCTGCCTTGAAAGGGGAC AACAGTGGAAACGACTCTAATACCGCATGAC GCATAGGGTGCATGATTTTATGCCAAAGAT TTATCGCTAAAGATGGCTCGCTGATTAG CTAGTTGTTGGGTAACGGCCACCAAGCGGA CGATCAGACGGACTGAAGAGTTGACCGC CACATGGACTGAGATACGCCAGACTCTA CGGGGAGCAGCTGGGGAAATATGGGAATG GGCGCAAGCTGACCAGAACCGCGGTGAA GGAAGAAGGCTTGGTTGAAACCTCTTTA AGAGGGAAGAGCAGAACAGCGTACCTCTAGAA TAAGCACCGCTAATACGTG	TGCTTAACACATGCAAGTCGAACGAGAACATGCTG AAGGAGGATTCTGCTCAAACCGAAGTAGAGGAAGT GGCGGAGGGTGAAGTAACGCGTGAGGAACCTGCC TTGAAGAGGGGACACAGTTGAAACAGACTGCT AAATACCGCATACGCGATAGGGGTCGCATGATCTT ATGCCAAAGATTATTGCTTCAAGATGGCTCGCG TCTGATTAGCTGGTTGGCGGGTAACGGGCCACCA AGGGCACGATCAGTAGCCGACTGAGAGGTTGAA CGGGCACATTGGGACTGAGATACGGCCAGACTCC TACGGGAGGCGCAGACTGGGGAAATGGCAATG GGCGCAAGCTGACCAGAACCGCGGTGAAAG AAAGAAGCTTCTGGGTGTAACCTTTAAAGAG GGAAGAGCAGAACAGCGTACCTCTAGAAATAAGCC ACGGCTAATACGTG
consensusSequence	Root(100);Bacteria(100);Firmicutes(100);Clos tridia(100);Clostridiales(100);Ruminococcace ae(100);Oscillibacter(99)	Root(100);Bacteria(100);Firmicutes(100);Clostri dia(100);Clostridiales(100);Ruminococcaceae(10 0);Oscillibacter(94)
rdpSummary		
UCHIMERefScore	0.038	0.0612
UCHIMEDenovoScore	0	0.1856
maxUCHIMEScore	0.038	0.1856
UCHIMEVerdict	neither	neither
toGoldGlobalPercentIdentity	90.59	88.44
toGoldGlobalBestHit	249174	32020
toGoldHumanGlobalPercentIdentity	87.29	88.44

toGoldHumanGlobalBestHit	804634	32020
toHMPStrainsGlobalPercentIdentity	87.29	88.44
toHmpStrainsGlobalBestHit	804634	32020
toSilvaGlobalPercentIdentity	98.95	99.78
toSilvaGlobalBestHit	Unl08zxz	Unl022fr
toGreengenesNamedGlobalPercentIdentity	92.45	91.61
toGreengenesNamedGlobalBestHit	607206	607206
toGreengenesUnnamedGlobalPercentIdentity	91.08	91.16
toGreengenesUnnamedGlobalBestHit	244027	812374
maxFraction	0.6979866	0.3489933
maxFractionBodyHabitat	Stool	Stool
454_seqCounts_Stool	13561	709
454_seqCounts_Buccal mucosa	8	3
454_seqCounts_Hard palate	37	1
454_seqCounts_Keratinized gingiva	3	0
454_seqCounts_Palatine Tonsils	6	0
454_seqCounts_Saliva	11	0
454_seqCounts_Subgingival plaque	0	0
454_seqCounts_Supragingival plaque	0	0
454_seqCounts_Throat	19	1
454_seqCounts_Tongue dorsum	0	2
454_seqCounts_Anterior nares	161	6
454_seqCounts_L_Antecubital fossa	364	15
454_seqCounts_L_Retroauricular crease	112	8
454_seqCounts_R_Antecubital fossa	341	37
454_seqCounts_R_Retroauricular crease	204	6
454_seqCounts_Mid vagina	7	0
454_seqCounts_Posterior fornix	10	0
454_seqCounts_Vaginal introitus	44	0
454_seqCounts_positive	0	0
454_seqCounts_water	0	0
454_subjectCounts_Stool	104	52
454_subjectCounts_Buccal mucosa	5	2
454_subjectCounts_Hard palate	9	1

454_subjectCounts_Keratinized gingiva	3	0	454_subjectfractions_Vaginal introitus	0.13235295	0
454_subjectCounts_Palatine Tonsils	4	0	454_subjectfractions_positive	0	0
454_subjectCounts_Saliva	6	0	454_subjectfractions_water	0	0
454_subjectCounts_Subgingival plaque	0	0	454_RelativeAbundanceStool	0.007333881	3.07E-04
454_subjectCounts_Supragingival plaque	0	0	454_RelativeAbundanceBuccal mucosa	0.00000398	9.16E-07
454_subjectCounts_Throat	3	1	454_RelativeAbundanceHard palate	0.000095	9.62E-07
454_subjectCounts_Tongue dorsum	0	1	454_RelativeAbundanceKeratinized gingiva	0.00000379	0
454_subjectCounts_Anterior nares	18	3	454_RelativeAbundancePalatine Tonsils	0.00000542	0
454_subjectCounts_L_Antecubital fossa	22	6	454_RelativeAbundanceSaliva	0.0000109	0
454_subjectCounts_L_Retroauricular crease	13	3	454_RelativeAbundanceSubgingival plaque	0	0
454_subjectCounts_R_Antecubital fossa	21	5	454_RelativeAbundanceSupragingival plaque	0	0
454_subjectCounts_R_Retroauricular crease	11	1	454_RelativeAbundanceThroat	0.0000492	7.04E-07
454_subjectCounts_Mid vagina	2	0	454_RelativeAbundanceTongue dorsum	0	8.48E-07
454_subjectCounts_Posterior fornix	2	0	454_RelativeAbundanceAnterior nares	0.00013	5.07E-06
454_subjectCounts_Vaginal introitus	9	0	454_RelativeAbundanceL_Antecubital fossa	0.000338	8.00E-06
454_subjectCounts_positive	0	0	454_RelativeAbundanceL_Retroauricular crease	0.000153	4.02E-06
454_subjectCounts_water	0	0	454_RelativeAbundanceR_Antecubital fossa	0.000878	3.11E-05
454_subjectfractions_Stool	0.6979866	0.3489933	454_RelativeAbundanceR_Retroauricular crease	0.0001	9.22E-07
454_subjectfractions_Buccal mucosa	0.035714287	0.014285714	454_RelativeAbundanceMid vagina	0.00000316	0
454_subjectfractions_Hard palate	0.06666667	0.007407407	454_RelativeAbundancePosterior fornix	0.00000454	0
454_subjectfractions_Keratinized gingiva	0.021276595	0	454_RelativeAbundanceVaginal introitus	0.0000574	0
454_subjectfractions_Palatine Tonsils	0.028368793	0	454_RelativeAbundancepositive	0	0
454_subjectfractions_Saliva	0.045801528	0	454_RelativeAbundancewater	0	0
454_subjectfractions_Subgingival plaque	0	0			
454_subjectfractions_Supragingival plaque	0	0			
454_subjectfractions_Throat	0.022222223	0.007407407			
454_subjectfractions_Tongue dorsum	0	0.006896552			
454_subjectfractions_Anterior nares	0.1294964	0.021582734			
454_subjectfractions_L_Antecubital fossa	0.16058394	0.04379562			
454_subjectfractions_L_Retroauricular crease	0.084415585	0.019480519			
454_subjectfractions_R_Antecubital fossa	0.15	0.035714287			
454_subjectfractions_R_Retroauricular crease	0.07096774	0.006451613			
454_subjectfractions_Mid vagina	0.02739726	0			
454_subjectfractions_Posterior fornix	0.028169014	0			

Table S2

searching for 16S rRNA gene of
Caecococcus microfluidicus isolated in
this paper in nr database by BLAST
Database: nr
Fields: subject ids, % identity,
alignment length, mismatches, gap
opens, q. start, q. end, s. start, s. end,
evalue, bit score

#apply filter: percent identity greater or equal to 97%, and alignment length greater or equal to 250

gi 319500066 gb HQ792927.1	99.86	1453	2	0	1	1453	1	1453	0	2673
gi 261261403 gb GQ897250.1	99.72	1453	4	0	1	1453	19	1471	0	2662
gi 261261244 gb GQ897091.1	99.72	1453	4	0	1	1453	19	1471	0	2662
gi 126112143 gb EF041832.1	99.66	1453	5	0	1	1453	21	1473	0	2656
gi 126111973 gb EF041662.1	99.66	1453	5	0	1	1453	21	1473	0	2656
gi 126111971 gb EF041660.1	99.66	1453	5	0	1	1453	21	1473	0	2656
gi 126111482 gb EF041171.1	99.66	1453	5	0	1	1453	21	1473	0	2656
gi 126111445 gb EF041134.1	99.66	1453	5	0	1	1453	21	1473	0	2656
gi 126111306 gb EF400995.1	99.66	1453	5	0	1	1453	21	1473	0	2656
gi 126111276 gb EF400965.1	99.66	1453	5	0	1	1453	21	1473	0	2656
gi 126111154 gb EF400843.1	99.66	1453	5	0	1	1453	21	1473	0	2656
gi 261261148 gb GQ896995.1	99.59	1453	6	0	1	1453	19	1471	0	2651
gi 126112165 gb EF041854.1	99.59	1453	6	0	1	1453	21	1473	0	2651
gi 126111704 gb EF041393.1	99.59	1453	6	0	1	1453	21	1473	0	2651
gi 126111580 gb EF401269.1	99.59	1453	6	0	1	1453	21	1473	0	2651
gi 126111559 gb EF041248.1	99.59	1453	6	0	1	1453	21	1473	0	2651
gi 126111536 gb EF041225.1	99.59	1453	6	0	1	1453	21	1473	0	2651
gi 126111075 gb EF400764.1	99.59	1453	6	0	1	1453	21	1473	0	2651
gi 319493440 gb HQ786301.1	99.52	1453	7	0	1	1453	1	1453	0	2645
gi 261261697 gb GQ897544.1	99.52	1453	7	0	1	1453	19	1471	0	2645
gi 261261684 gb GQ897531.1	99.52	1453	7	0	1	1453	19	1471	0	2645
gi 261261500 gb GQ897347.1	99.52	1453	7	0	1	1453	19	1471	0	2645
gi 261261113 gb GQ896960.1	99.52	1453	7	0	1	1453	19	1471	0	2645
gi 126115125 gb EF404805.1	99.52	1453	7	0	1	1453	22	1474	0	2645
gi 126111760 gb EF401449.1	99.52	1453	7	0	1	1453	21	1473	0	2645
gi 126111328 gb EF401017.1	99.52	1453	7	0	1	1453	21	1473	0	2645
gi 126111235 gb EF400924.1	99.52	1453	7	0	1	1453	21	1473	0	2645
gi 126111138 gb EF400827.1	99.52	1453	7	0	1	1453	21	1473	0	2645
gi 319495953 gb HQ788814.1	99.93	1434	1	0	20	1453	1	1434	0	2643
gi 126111646 gb EF401335.1	99.52	1453	6	1	1	1453	21	1472	0	2643
gi 126111403 gb EF401092.1	99.52	1453	6	1	1	1453	21	1472	0	2643
gi 126111287 gb EF400976.1	99.52	1453	6	1	1	1453	21	1472	0	2643
gi 261262538 gb GQ898389.1	99.45	1453	8	0	1	1453	20	1472	0	2639
gi 261262369 gb GQ898220.1	99.45	1453	8	0	1	1453	20	1472	0	2639
gi 126115648 gb EF405325.1	99.45	1453	8	0	1	1453	22	1474	0	2639
gi 126115366 gb EF405043.1	99.45	1453	8	0	1	1453	22	1474	0	2639
gi 126115362 gb EF405039.1	99.45	1453	8	0	1	1453	22	1474	0	2639
gi 126111838 gb EF401257.1	99.45	1453	8	0	1	1453	21	1473	0	2639
gi 6456061 gb AF132255.1	99.52	1452	3	4	4	1453	24	1473	0	2639
gi 319482775 gb HQ775636.1	99.52	1449	7	0	5	1453	1	1449	0	2638
gi 319499423 gb HQ792284.1	99.45	1450	8	0	2	1451	1	1450	0	2634
gi 319488021 gb HQ780882.1	99.65	1441	5	0	4	1444	3	1443	0	2634
gi 261262337 gb GQ898188.1	99.38	1453	9	0	1	1453	20	1472	0	2634
gi 261261861 gb GQ897712.1	99.38	1453	9	0	1	1453	19	1471	0	2634
gi 126115512 gb EF405189.1	99.38	1453	9	0	1	1453	22	1474	0	2634
gi 126115207 gb EF404884.1	99.38	1453	9	0	1	1453	22	1474	0	2634
gi 126112029 gb EF401718.1	99.38	1453	9	0	1	1453	21	1473	0	2634
gi 126111899 gb EF401588.1	99.38	1453	9	0	1	1453	21	1473	0	2634
gi 126111893 gb EF401582.1	99.38	1453	9	0	1	1453	21	1473	0	2634
gi 126111713 gb EF401402.1	99.38	1453	9	0	1	1453	21	1473	0	2634
gi 126111061 gb EF400750.1	99.38	1453	9	0	1	1453	21	1473	0	2634
gi 319487719 gb HQ780580.1	99.38	1450	9	0	4	1453	1	1450	0	2628
gi 261262554 gb GQ898405.1	99.31	1453	10	0	1	1453	20	1472	0	2628
gi 126112132 gb EF401821.1	99.31	1453	10	0	1	1453	21	1473	0	2628
gi 126112111 gb EF401800.1	99.31	1453	10	0	1	1453	21	1473	0	2628
gi 126112073 gb EF401762.1	99.31	1453	10	0	1	1453	21	1473	0	2628
gi 126112033 gb EF401722.1	99.31	1453	10	0	1	1453	21	1473	0	2628

g 1261112002 gb EF401691.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111979 gb EF401668.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111925 gb EF401614.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111813 gb EF401502.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111660 gb EF401349.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111649 gb EF401338.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111596 gb EF401285.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111542 gb EF401231.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111535 gb EF401224.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111465 gb EF401154.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111399 gb EF401088.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111378 gb EF401067.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111376 gb EF401065.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111363 gb EF401052.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111353 gb EF401042.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111323 gb EF401012.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111201 gb EF400890.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111135 gb EF400824.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 126111085 gb EF400774.1	99.31	1453	10	0	1	1453	21	1473	0	2628
g 319490773 gb HQ783634.1	99.31	1450	10	0	4	1453	1	1450	0	2623
g 261261792 gb GQ897639.1	99.24	1453	11	0	1	1453	19	1471	0	2623
g 126115092 gb EF404772.1	99.24	1453	11	0	1	1453	22	1474	0	2623
g 126114933 gb EF404613.1	99.24	1453	11	0	1	1453	22	1474	0	2623
g 126112161 gb EF401850.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126112160 gb EF401849.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126112061 gb EF401750.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111968 gb EF401657.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111930 gb EF401619.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111902 gb EF401591.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111879 gb EF401568.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111753 gb EF401442.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111737 gb EF401426.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111613 gb EF401302.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111405 gb EF401094.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 126111362 gb EF401051.1	99.24	1453	11	0	1	1453	21	1473	0	2623
g 319488801 gb HQ781662.1	99.24	1452	10	1	2	1452	1	1452	0	2619
g 319499925 gb HQ792786.1	99.44	1441	8	0	13	1453	1	1441	0	2617
g 319492434 gb HQ785295.1	99.17	1454	11	1	1	1453	2	1455	0	2617
g 319488912 gb HQ781773.1	99.24	1450	11	0	4	1453	1	1450	0	2617
g 261262501 gb GQ898352.1	99.17	1451	12	0	3	1453	20	1470	0	2617
g 261260989 gb GQ896836.1	99.17	1453	12	0	1	1453	19	1471	0	2617
g 126111846 gb EF401535.1	99.17	1453	12	0	1	1453	21	1473	0	2617
g 126111498 gb EF401187.1	99.17	1453	12	0	1	1453	21	1473	0	2617
g 126111494 gb EF401183.1	99.17	1453	12	0	1	1453	21	1473	0	2617
g 126111063 gb EF400752.1	99.17	1453	12	0	1	1453	21	1473	0	2617
g 1261115692 gb EF405369.1	99.17	1453	11	1	1	1453	22	1473	0	2615
g 319484767 gb HQ777628.1	99.38	1442	9	0	12	1453	1	1442	0	2614
g 319494580 gb HQ787441.1	99.17	1450	12	0	4	1453	1	1450	0	2612
g 261261902 gb GQ897753.1	99.11	1453	13	0	1	1453	19	1471	0	2612
g 261261832 gb GQ897683.1	99.11	1453	13	0	1	1453	18	1470	0	2612
g 126111936 gb EF401625.1	99.11	1453	13	0	1	1453	21	1473	0	2612
g 126111726 gb EF401451.1	99.11	1453	13	0	1	1453	21	1473	0	2612
g 126111600 gb EF401289.1	99.11	1453	13	0	1	1453	21	1473	0	2612
g 126111231 gb EF400920.1	99.11	1453	13	0	1	1453	21	1473	0	2612
g 18644507 gb AF371798.1	99.58	1431	4	2	1	1430	21	1450	0	2612
g 319494186 gb HQ787047.1	99.17	1448	12	0	4	1451	1	1448	0	2608
g 319493693 gb HQ786554.1	99.1	1451	13	0	3	1453	1	1451	0	2608
g 240005432 gb GQ158590.1	99.17	1448	12	0	6	1453	1	1448	0	2608
g 319499385 gb HQ792246.1	99.37	1439	8	1	9	1446	1	1439	0	2606
g 319494484 gb HQ787345.1	99.24	1444	11	0	1	1444	5	1448	0	2606
g 319499757 gb HQ792618.1	99.24	1443	11	0	5	1447	1	1443	0	2604
g 319466460 gb HQ759321.1	99.24	1440	11	0	14	1453	1	1440	0	2601
g 319500228 gb HQ793089.1	99.17	1444	12	0	7	1450	1	1444	0	2601
g 319500039 gb HQ792900.1	99.37	1435	9	0	1	1435	3	1437	0	2601
g 319494363 gb HQ787224.1	99.03	1450	14	0	4	1453	1	1450	0	2601
g 1261115689 gb EF405366.1	98.97	1453	15	0	1	1453	22	1474	0	2601
g 1261115469 gb EF405146.1	98.97	1453	15	0	1	1453	22	1474	0	2601
g 1261115408 gb EF405085.1	98.97	1453	15	0	1	1453	22	1474	0	2601

gi 319517965 gb HQ810826.1	99.03	1449	14	0	3	1451	1	1449	0	2599		gi 219533792 gb FJ510597.1	99.63	1364	5	0	2	1365	13	1376	0	2492
gi 319494413 gb HQ787274.1	99.17	1442	12	0	12	1453	1	1442	0	2597		gi 31949960 gb HQ792821.1	97.65	1448	34	0	4	1451	1	1448	0	2486
gi 319498028 gb HQ781889.1	99.1	1444	13	0	10	1453	1	1444	0	2595		gi 110446177 gb DQ806392.1	99.42	1370	6	2	1	1368	33	1402	0	2484
gi 240006081 gb GQ159239.1	98.9	1453	16	0	1	1453	1	1453	0	2595		gi 388933029 gb JQ188146.1	99.85	1350	2	0	1	1350	1	1350	0	2483
gi 126115476 gb FE405153.1	98.9	1453	16	0	1	1453	22	1474	0	2595		gi 388933200 gb JQ188317.1	99.85	1350	2	0	1	1350	1	1350	0	2483
gi 126115363 gb FE405040.1	98.9	1453	16	0	1	1453	22	1474	0	2595		gi 388932914 gb JQ188031.1	99.85	1350	2	0	1	1350	1	1350	0	2483
gi 319495888 gb HQ788749.1	99.24	1438	10	1	17	1453	1	1438	0	2593		gi 110441168 gb DQ800768.1	99.42	1368	8	0	1	1368	33	1400	0	2483
gi 319494567 gb HQ787428.1	99.1	1443	13	0	11	1453	1	1443	0	2593		gi 319466572 gb HQ759433.1	98.71	1397	18	0	5	1401	1	1397	0	2481
gi 319494832 gb HQ787693.1	99.17	1439	12	0	1	1439	8	1446	0	2591		gi 219534086 gb FI510891.1	99.49	1363	7	0	1	1363	10	1372	0	2479
gi 319466750 gb HQ759611.1	99.37	1430	9	0	19	1448	1	1430	0	2591		gi 388928035 gb JQ183152.1	99.78	1350	3	0	1	1350	1	1350	0	2477
gi 319478696 gb HQ780557.1	99.03	1444	14	0	1	1444	7	1450	0	2590		gi 388935405 gb JQ190522.1	99.78	1350	3	0	1	1350	1	1350	0	2477
gi 240006082 gb GQ159240.1	98.83	1453	17	0	1	1453	1	1453	0	2590		gi 388933350 gb JQ188467.1	99.78	1350	3	0	1	1350	1	1350	0	2477
gi 240006079 gb GQ159237.1	98.83	1454	15	2	1	1453	1	1453	0	2590		gi 388933228 gb JQ188345.1	99.78	1350	3	0	1	1350	1	1350	0	2477
gi 319493543 gb HQ78640.1	99.1	1440	13	0	14	1453	1	1440	0	2588		gi 110449771 gb DQ800986.1	99.34	1368	9	0	1	1368	32	1399	0	2477
gi 319488000 gb HQ780861.1	99.03	1443	14	0	1	1443	10	1452	0	2588		gi 110449667 gb DQ800982.1	99.34	1368	9	0	1	1368	33	1400	0	2477
gi 240006083 gb GQ159241.1	98.83	1453	16	1	1	1453	1	1452	0	2588		gi 110448825 gb DQ800940.1	99.34	1368	9	0	1	1368	21	1388	0	2477
gi 170722519 gb EU513967.1	99.17	1438	10	2	1	1438	1	1436	0	2588		gi 110436401 gb DQ796001.1	99.34	1368	9	0	1	1368	21	1388	0	2477
gi 319496000 gb HQ788861.1	99.16	1436	12	0	18	1453	1	1436	0	2586		gi 388933272 gb JQ188389.1	99.7	1352	2	0	1	1350	1	1352	0	2473
gi 319494044 gb HQ786905.1	99.03	1441	14	0	1	1441	4	1444	0	2584		gi 219531115 gb FI507920.1	99.63	1354	5	0	12	1365	1	1354	0	2473
gi 319478727 gb HQ780588.1	98.89	1447	16	0	2	1448	1	1447	0	2584		gi 388931523 gb JQ186640.1	99.7	1350	4	0	1	1350	1	1350	0	2471
gi 319485110 gb HQ777971.1	98.96	1444	15	0	10	1453	1	1444	0	2584		gi 388935595 gb JQ190712.1	99.7	1350	4	0	1	1350	1	1350	0	2471
gi 319494343 gb HQ742204.1	99.1	1438	13	0	1	1438	1	1438	0	2584		gi 388935367 gb JQ190484.1	99.7	1350	4	0	1	1350	1	1350	0	2471
gi 324959751 gb HQ716104.1	98.76	1453	18	0	1	1453	19	1471	0	2584		gi 388935044 gb JQ190161.1	99.7	1350	4	0	1	1350	1	1350	0	2471
gi 240006080 gb GQ159238.1	98.76	1454	15	3	1	1453	1	1452	0	2582		gi 219536114 gb FI512919.1	99.34	1365	9	0	1	1365	11	1375	0	2471
gi 319517807 gb HQ810668.1	98.96	1437	15	0	17	1453	1	1437	0	2571		gi 219531116 gb FI507921.1	99.34	1365	9	0	1	1365	11	1375	0	2471
gi 319494034 gb HQ786895.1	98.96	1436	15	0	18	1453	1	1436	0	2569		gi 192988166 gb EU778163.1	99.27	1368	10	0	1	1368	33	1400	0	2471
gi 319499806 gb HQ792667.1	98.75	1443	18	0	11	1453	1	1443	0	2566		gi 192981066 gb EU775089.1	99.27	1368	10	0	1	1368	33	1400	0	2471
gi 319495841 gb HQ788702.1	98.88	1434	16	0	18	1451	1	1434	0	2560		gi 192976430 gb EU772453.1	99.27	1368	10	0	1	1368	33	1400	0	2471
gi 319466752 gb HQ759613.1	98.54	1441	20	1	4	1443	1	1441	0	2543		gi 169278176 gb E462701.1	99.27	1368	10	0	1	1368	21	1388	0	2471
gi 319466850 gb HQ759711.1	98.33	1440	24	0	12	1451	1	1440	0	2577		gi 169278092 gb E462617.1	99.27	1368	10	0	1	1368	21	1388	0	2471
gi 319487226 gb HQ780587.1	98.33	1436	22	2	1	1436	3	1436	0	2518		gi 110441305 gb DQ800905.1	99.27	1368	10	0	1	1368	33	1400	0	2471
gi 319466599 gb HQ759460.1	98.52	1423	19	2	12	1434	2	1422	0	2510		gi 110436243 gb DQ795843.1	99.27	1368	10	0	1	1368	33	1400	0	2471
gi 319500123 gb HQ792984.1	97.99	1445	28	1	1	1445	6	1449	0	2507		gi 388933965 gb JQ189082.1	99.7	1350	3	1	1	1350	1	1349	0	2470
gi 110446538 gb DQ806753.1	99.56	1368	6	0	1	1368	32	1399	0	2494		gi 219531114 gb FI507919.1	99.34	1365	8	1	1	1365	11	1374	0	2470

g 388928800 gb JQ183917.1	99.63	1350	5	0	1	1350	1	1350	0	2466		g 62760514 gb AY981024.1	99.48	1350	7	0	1	1350	1	1350	0	2455
g 388935165 gb JQ190282.1	99.63	1350	5	0	1	1350	1	1350	0	2466		g 388935424 gb JQ190541.1	99.41	1354	4	4	1	1350	1	1354	0	2453
g 388934187 gb JQ189304.1	99.63	1350	5	0	1	1350	1	1350	0	2466		g 219536110 gb FJ512915.1	99.12	1364	12	0	2	1365	13	1376	0	2453
g 219536113 gb FJ512918.1	99.27	1365	10	0	1	1365	11	1375	0	2466		g 219535544 gb FJ512349.1	99.12	1364	12	0	2	1365	11	1374	0	2453
g 192981059 gb E775081.1	99.2	1368	11	0	1	1368	33	1400	0	2466		g 21953017 gb FJ509822.1	99.12	1364	12	0	1	1364	11	1374	0	2453
g 110446130 gb DQ806345.1	99.2	1368	11	0	1	1368	4	1371	0	2466		g 219533015 gb FJ509820.1	99.19	1361	11	0	4	1364	14	1374	0	2453
g 62764679 gb AY985189.1	99.63	1350	5	0	1	1350	1	1350	0	2466		g 388930479 gb JQ185596.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 219533434 gb FJ510146.1	99.27	1364	10	0	1	1364	11	1374	0	2464		g 388930251 gb JQ185368.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 110449111 gb DQ809326.1	99.2	1368	10	1	1	1368	33	1399	0	2464		g 388928126 gb JQ183243.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 388931569 gb JQ186686.1	99.56	1352	4	2	1	1350	1	1352	0	2462		g 388934631 gb JQ189748.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 388931221 gb JQ186338.1	99.56	1350	6	0	1	1350	1	1350	0	2460		g 388934877 gb JQ189994.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 388930259 gb JQ185376.1	99.56	1350	6	0	1	1350	1	1350	0	2460		g 388934922 gb JQ190039.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 388928405 gb JQ183522.1	99.56	1350	6	0	1	1350	1	1350	0	2460		g 388934445 gb JQ189562.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 388934145 gb JQ189262.1	99.56	1350	6	0	1	1350	1	1350	0	2460		g 388933151 gb JQ188268.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 388933765 gb JQ188882.1	99.56	1350	6	0	1	1350	1	1350	0	2460		g 322160786 gb JF175380.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 219536112 gb FJ512917.1	99.19	1365	11	0	1	1365	11	1375	0	2460		g 219535545 gb FJ512350.1	99.12	1363	11	1	3	1365	12	1373	0	2449
g 219536107 gb FJ512912.1	99.19	1365	11	0	1	1365	11	1375	0	2460		g 219533789 gb FJ510594.1	99.05	1366	11	2	1	1365	11	1375	0	2449
g 219536108 gb FJ512913.1	99.19	1365	11	0	1	1365	11	1375	0	2460		g 219533342 gb FJ510147.1	99.12	1362	12	0	3	1364	13	1374	0	2449
g 219533791 gb FJ510596.1	99.19	1365	11	0	1	1365	11	1375	0	2460		g 214018748 gb FJ364361.1	99.26	1356	10	0	1	1356	21	1376	0	2449
g 192976454 gb E772477.1	99.12	1368	12	0	1	1368	33	1400	0	2460		g 169291316 gb EU475841.1	98.98	1368	14	0	1	1368	21	1388	0	2449
g 169278049 gb EU462574.1	99.12	1368	12	0	1	1368	21	1388	0	2460		g 110436585 gb DQ796185.1	98.98	1368	14	0	1	1368	21	1388	0	2449
g 110449321 gb DQ809536.1	99.12	1368	12	0	1	1368	33	1400	0	2460		g 62764715 gb AY985225.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 219533340 gb FJ510145.1	99.19	1364	11	0	1	1364	11	1374	0	2459		g 62763312 gb AY983822.1	99.41	1350	8	0	1	1350	1	1350	0	2449
g 214017356 gb FJ362969.1	99.41	1355	8	0	1	1355	1	1355	0	2459		g 388933707 gb JQ188824.1	99.41	1350	6	2	1	1348	1	1350	0	2447
g 219533343 gb FJ510148.1	99.26	1360	10	0	5	1364	15	1374	0	2457		g 219533339 gb FJ510144.1	99.19	1358	11	0	7	1364	6	1363	0	2447
g 169276654 gb E461179.1	99.05	1370	11	2	1	1368	21	1390	0	2457		g 219533338 gb FJ510143.1	99.05	1364	13	0	1	1364	11	1374	0	2447
g 388929811 gb JQ184928.1	99.48	1351	6	1	1	1350	1	1351	0	2455		g 110436110 gb DQ795710.1	99.05	1364	13	0	1	1364	33	1396	0	2447
g 388928407 gb JQ183524.1	99.48	1350	7	0	1	1350	1	1350	0	2455		g 388934590 gb JQ189707.1	99.33	1352	7	2	1	1350	1	1352	0	2446
g 388935013 gb JQ190130.1	99.48	1350	7	0	1	1350	1	1350	0	2455		g 219531117 gb FJ507922.1	99.26	1354	10	0	12	1365	1	1354	0	2446
g 388935324 gb JQ190441.1	99.48	1350	7	0	1	1350	1	1350	0	2455		g 388934888 gb JQ190005.1	99.26	1356	4	6	1	1350	1	1356	0	2444
g 388932549 gb JQ187666.1	99.48	1351	6	1	1	1350	1	1351	0	2455		g 388934140 gb JQ189257.1	99.33	1351	8	1	1	1350	1	1351	0	2444
g 219536109 gb FJ512911.1	99.12	1365	12	0	1	1365	11	1375	0	2455		g 388933499 gb JQ188616.1	99.33	1350	9	0	1	1350	1	1350	0	2444
g 192981007 gb E775030.1	99.05	1368	13	0	1	1368	25	1392	0	2455		g 322160776 gb JF175370.1	99.33	1350	9	0	1	1350	1	1350	0	2444
g 110448398 gb DQ808613.1	99.05	1368	13	0	1	1368	21	1388	0	2455		g 219535543 gb FJ512348.1	98.97	1365	14	0	1	1365	11	1375	0	2444

gi 219531288 gb FJ508093.1	99.33	1350	9	0	1	1350	16	1365	0	2444
gi 219531287 gb FJ508092.1	99.33	1350	9	0	1	1350	16	1365	0	2444
gi 219531286 gb FJ508091.1	99.33	1350	9	0	1	1350	16	1365	0	2444
gi 219531284 gb FJ508089.1	99.33	1350	9	0	1	1350	16	1365	0	2444
gi 219531285 gb FJ508090.1	99.33	1350	9	0	1	1350	16	1365	0	2444
gi 214022757 gb FJ368373.1	99.19	1356	11	0	1	1356	1	1356	0	2444
gi 214018691 gb FJ364304.1	99.19	1356	11	0	1	1356	1	1356	0	2444
gi 192968905 gb EU764690.1	99.33	1350	9	0	1	1350	1	1350	0	2444
gi 62765741 gb AY986251.1	99.33	1350	9	0	1	1350	1	1350	0	2444
gi 62764956 gb AY985466.1	99.33	1350	9	0	1	1350	1	1350	0	2444
gi 219536109 gb FJ512914.1	98.97	1364	14	0	2	1365	13	1376	0	2442
gi 219536105 gb FJ512910.1	98.97	1365	13	1	1	1365	11	1374	0	2442
gi 219531289 gb FJ508094.1	99.41	1346	8	0	1	1346	16	1361	0	2442
gi 219531118 gb FJ507923.1	99.19	1355	11	0	11	1365	1	1355	0	2442
gi 388928236 gb JQ183353.1	99.26	1352	8	2	1	1350	1	1352	0	2440
gi 388934843 gb JQ189960.1	99.26	1352	8	2	1	1350	1	1352	0	2440
gi 388933452 gb JQ188569.1	99.26	1353	7	3	1	1350	1	1353	0	2440
gi 62765667 gb AY986177.1	99.26	1352	8	2	1	1350	1	1352	0	2440
gi 388931911 gb JQ187028.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388931775 gb JQ186892.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388931727 gb JQ186844.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388930236 gb JQ185353.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388928164 gb JQ183281.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388928479 gb JQ183596.1	99.26	1351	9	1	1	1350	1	1351	0	2438
gi 388928666 gb JQ183783.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388935278 gb JQ190395.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388934715 gb JQ189832.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388934682 gb JQ189799.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388933162 gb JQ188279.1	99.33	1347	9	0	4	1350	4	1350	0	2438
gi 388933068 gb JQ188185.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 388930723 gb JQ185840.1 ;gi 388	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 219535542 gb FJ512347.1	98.9	1365	15	0	1	1365	11	1375	0	2438
gi 219533788 gb FJ510593.1	98.9	1365	15	0	1	1365	11	1375	0	2438
gi 219531276 gb FJ508081.1	99.26	1350	10	0	1	1350	16	1365	0	2438
gi 219531277 gb FJ508082.1 ;gi 219	99.26	1350	10	0	1	1350	16	1365	0	2438
gi 214017889 gb FJ363502.1	99.12	1356	12	0	1	1356	1	1356	0	2438
gi 214017490 gb FJ363103.1	99.12	1356	12	0	1	1356	1	1356	0	2438
gi 192970039 gb EU765824.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 192968817 gb EU764602.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 192968594 gb EU764379.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 192981510 gb EU775533.1	98.83	1368	16	0	1	1368	33	1400	0	2438
gi 169285159 gb EU469684.1	98.83	1368	16	0	1	1368	32	1399	0	2438
gi 62759235 gb AY979745.1 ;gi 388	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 62765196 gb AY985706.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 62765128 gb AY985638.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 62765091 gb AY985601.1	99.26	1351	8	2	1	1350	1	1350	0	2438
gi 62764908 gb AY985418.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 62764829 gb AY985339.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 62763469 gb AY983979.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 62763277 gb AY983787.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 62757651 gb AY978161.1	99.26	1350	10	0	1	1350	1	1350	0	2438
gi 62757223 gb AY97733.1	99.26	1351	8	2	1	1350	1	1350	0	2438
gi 322160944 gb JF175538.1	99.26	1350	9	1	1	1350	1	1349	0	2436
gi 322153252 gb JF167846.1	99.26	1350	9	1	1	1350	1	1349	0	2436
gi 219533790 gb FJ510595.1	98.9	1365	14	1	1	1365	11	1374	0	2436
gi 219531275 gb FJ508080.1	99.26	1349	10	0	2	1350	17	1365	0	2436
gi 214018920 gb FJ364534.1	98.83	1367	16	0	1	1367	1	1367	0	2436
gi 388931650 gb JQ186767.1	99.19	1352	9	2	1	1350	1	1352	0	2435
gi 219533444 gb FJ510149.1	98.97	1360	14	0	5	1364	15	1374	0	2435
gi 219533016 gb FJ509821.1	99.04	1357	13	0	8	1364	17	1373	0	2435
gi 219532157 gb FJ508962.1	98.9	1363	14	1	1	1363	11	1372	0	2435
gi 62765608 gb AY986118.1	99.19	1353	8	3	1	1350	1	1353	0	2435
gi 62765449 gb AY985959.1	99.19	1352	9	2	1	1350	1	1352	0	2435
gi 388931501 gb JQ186618.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 388928698 gb JQ183815.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 388928240 gb JQ183357.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 388935772 gb JQ190889.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 388934425 gb JQ189542.1	99.19	1350	11	0	1	1350	1	1350	0	2433

gi 322146999 gb JF161593.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 219531283 gb FJ508088.1	99.19	1350	11	0	1	1350	16	1365	0	2433
gi 219531282 gb FJ508087.1	99.19	1350	11	0	1	1350	16	1365	0	2433
gi 214021126 gb FJ366742.1	99.26	1348	9	1	10	1356	35	1382	0	2433
gi 214020057 gb FJ365672.1	99.26	1347	10	0	10	1356	2	1348	0	2433
gi 192980034 gb EU774057.1	98.76	1368	17	0	1	1368	21	1388	0	2433
gi 62765226 gb AY985736.1 ;gi 1929	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 62765100 gb AY985610.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 62765074 gb AY985584.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 62764776 gb AY985286.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 62763468 gb AY983978.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 62763355 gb AY983865.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 62759806 gb AY980316.1 ;gi 3889	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 62757414 gb AY977924.1	99.19	1350	11	0	1	1350	1	1350	0	2433
gi 219531272 gb FJ508077.1	99.26	1346	10	0	1	1346	16	1361	0	2431
gi 219531273 gb FJ508078.1	99.26	1346	10	0	1	1346	16	1361	0	2431
gi 219531270 gb FJ508075.1	99.26	1346	10	0	1	1346	16	1361	0	2431
gi 214026068 gb FJ371687.1	99.04	1355	13	0	1	1355	1	1355	0	2431
gi 214018797 gb FJ364410.1	98.83	1364	16	0	1	1364	1	1364	0	2431
gi 62763276 gb AY983786.1	99.19	1350	10	1	1	1350	1	1349	0	2431
gi 388928008 gb JQ183125.1	99.11	1353	9	3	1	1350	1	1353	0	2429
gi 219531290 gb FJ508095.1	99.26	1345	10	0	1	1345	16	1360	0	2429
gi 219531280 gb FJ508085.1	99.26	1345	10	0	1	1345	16	1360	0	2429
gi 219531268 gb FJ508073.1	99.26	1345	10	0	1	1345	16	1360	0	2429
gi 388931810 gb JQ186927.1	99.11	1351	11	1	1	1350	1	1351	0	2427
gi 388931288 gb JQ186405.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 388928632 gb JQ183749.1	99.11	1351	10	2	1	1350	1	1350	0	2427
gi 388928637 gb JQ183754.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 388930665 gb JQ185782.1	99.11	1351	10	2	1	1350	1	1350	0	2427
gi 388928400 gb JQ183517.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 388935714 gb JQ180831.1	99.11	1354	5	6	2	1350	3	1354	0	2427
gi 388934827 gb JQ189944.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 388933351 gb JQ188468.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 319484570 gb H0777431.1	97.6	1417	32	2	19	1435	1	1415	0	2427
gi 322160977 gb JF175571.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 219535541 gb FJ512346.1	98.75	1365	17	0	1	1365	11	1375	0	2427
gi 214026245 gb FJ371864.1	98.97	1357	13	1	1	1356	1	1357	0	2427
gi 192970466 gb EU766251.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 192970348 gb U766133.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 192968537 gb EU764322.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 192966307 gb EU762092.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 192966162 gb U761947.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 169278050 gb U462575.1	98.68	1368	18	0	1	1368	21	1388	0	2427
gi 110433710 gb DQ793310.1	98.68	1368	18	0	1	1368	21	1388	0	2427
gi 62759181 gb AY979691.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 62765793 gb AY986303.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 62764702 gb AY985212.1	99.11	1350	12	0	1	1350	1	1350	0	2427
gi 388934664 gb JQ189781.1	99.04	1354	9	4	1	1350	1	1354	0	2425
gi 322147187 gb JF161781.1	99.11	1350	11	1	1	1350	1	1349	0	2425
gi 219531281 gb FJ508086.1	99.11	1350	11	1	1	1350	16	1364	0	2425
gi 219531274 gb FJ508079.1	99.18	1346	11	0	1	1346	16	1361	0	2425
gi 219531271 gb FJ508076.1	99.18	1346	11	0	1	1346	16	1361	0	2425
gi 219531269 gb FJ508074.1	99.18	1346	11	0	1	1346	16	1361	0	2425
gi 219531279 gb FJ508084.1	99.11	1349	10	2	1	1348	16	1363	0	2423
gi 62764920 gb AY985430.1	99.04	1353	10	3	1	1350	1	1353	0	2423
gi 388930615 gb JQ185732.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 388930442 gb JQ185559.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 388929030 gb JQ184147.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 388928543 gb JQ183660.1	99.04	1351	11	2	1	1350	1	1350	0	2422
gi 219533787 gb FJ510592.1	98.68	1366	16	2	1	1365	11	1375	0	2422
gi 214026073 gb FJ371692.1	98.89	1356	15	0	1	1356	1	1356	0	2422
gi 192970627 gb U766412.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 192970356 gb U766141.1	99.04	1351	12	1	1	1350	1	1351	0	2422
gi 192969098 gb U764883.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 192981375 gb U775398.1	98.61	1368	19	0	1	1368	32	1399	0	2422
gi 169287503 gb U472028.1	98.61	1368	19	0	1	1368	33	1400	0	2422
gi 110437813 gb DQ797413.1	98.61	1368	19	0	1	1368	21	1388	0	2422
gi 110437539 gb DQ797139.1	98.61	1368	19	0	1	1368	33	1400	0	2422

gi 62759154 gb AY979664.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 62759099 gb AY979609.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 62765862 gb AY986372.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 62765215 gb AY985725.1	99.04	1350	13	0	1	1350	1	1350	0	2422
gi 322147075 gb JF161669.1	99.04	1350	12	1	1	1350	1	1349	0	2420
gi 192967875 gb EU763660.1	98.97	1354	10	4	1	1350	1	1354	0	2420
gi 62764708 gb AY985218.1	98.97	1354	10	4	1	1350	1	1354	0	2420
gi 388934798 gb JQ189915.1	98.96	1352	12	2	1	1350	1	1352	0	2418
gi 192970605 gb EU766390.1	98.97	1353	11	3	1	1350	1	1353	0	2418
gi 192988214 gb EU778185.1	98.54	1371	17	3	1	1368	9	1379	0	2418
gi 110441542 gb DQ801142.1	98.54	1371	17	3	1	1368	9	1379	0	2418
gi 388930601 gb JQ185718.1	98.96	1351	12	2	1	1350	1	1350	0	2416
gi 388928546 gb JQ183663.1	98.96	1352	11	3	1	1350	1	1351	0	2416
gi 388928362 gb JQ183479.1	98.96	1350	14	0	1	1350	1	1350	0	2416
gi 388934972 gb JQ190089.1	98.96	1351	13	1	1	1350	1	1351	0	2416
gi 319498064 gb H0790925.1	98.4	1375	20	2	23	1396	1	1374	0	2416
gi 219531266 gb FJ508071.1	99.11	1344	12	0	1	1344	16	1359	0	2416
gi 214022030 gb FJ367646.1	98.82	1356	16	0	1	1356	1	1356	0	2416
gi 192970585 gb EU766370.1	99.04	1349	10	3	1	1346	1	1349	0	2416
gi 192970493 gb EU766278.1	98.96	1351	13	1	1	1350	1	1351	0	2416
gi 192966008 gb EU761793.1	98.96	1352	11	3	1	1350	1	1351	0	2416
gi 169285267 gb EU469792.1	98.54	1368	20	0	1	1368	33	1400	0	2416
gi 169278106 gb EU462631.1	98.54	1369	18	2	1	1368	21	1388	0	2416
gi 169278096 gb EU462621.1	98.54	1368	20	0	1	1368	21	1388	0	2416
gi 169276721 gb EU461246.1	98.54	1368	20	0	1	1368	33	1400	0	2416
gi 110445418 gb DQ805633.1	98.54	1369	19	1	1	1368	21	1389	0	2416
gi 214018755 gb FJ364368.1	98.82	1355	16	0	1	1355	1	1355	0	2414
gi 388932925 gb JQ188042.1	98.89	1352	13	2	1	1350	1	1352	0	2412
gi 169291281 gb EU475806.1	98.47	1370	19	2	1	1368	21	1390	0	2412
gi 388935011 gb JQ190128.1	98.89	1350	15	0	1	1350	1	1350	0	2410
gi 388933914 gb JQ189031.1	98.82	1355	11	5	1	1350	1	1355	0	2409
gi 388933284 gb JQ188401.1	98.82	1355	11	5	1	1350	1	1355	0	2409
gi 219536111 gb FJ512916.1	98.61	1364	14	5	3	1365	12	1371	0	2409
gi 192965883 gb EU761668.1	98.82	1355	11	5	1	1350	1	1355	0	2409
gi 110445082 gb DQ805297.1	98.39	1370	20	2	1	1368	4	1373	0	2407
gi 388935520 gb JQ190637.1	98.82	1351	15	1	1	1350	1	1351	0	2405
gi 388935228 gb JQ190345.1	98.81	1350	16	0	1	1350	1	1350	0	2405
gi 214026130 gb FJ371749.1	99.18	1335	11	0	1	1335	1	1335	0	2405
gi 214017608 gb FJ363221.1	98.67	1356	18	0	1	1356	1	1356	0	2405
gi 214017504 gb FJ363117.1	98.67	1356	18	0	1	1356	1	1356	0	2405
gi 192972226 gb EU768011.1	98.89	1348	13	2	1	1346	1	1348	0	2405
gi 169282771 gb EU467296.1	99.1	1338	12	0	14	1351	1	1338	0	2405
gi 219532455 gb FJ509260.1	98.6	1359	17	2	1	1359	11	1367	0	2403
gi 214026456 gb FJ372075.1	99.03	1340	13	0	17	1356	3	1342	0	2403
gi 192970718 gb EU766503.1	98.74	1354	13	4	1	1350	1	1354	0	2403
gi 192980909 gb EU774932.1	98.32	1368	23	0	1	1368	21	1388	0	2399
gi 192980191 gb EU774214.1	99.1	1335	12	0	1	1335	21	1355	0	2399
gi 388930741 gb JQ185858.1	98.67	1354	14	4	1	1350	1	1354	0	2398
gi 388930096 gb JQ185213.1	98.67	1355	13	4	1	1350	1	1355	0	2398
gi 388928799 gb JQ183916.1	98.67	1355	13	5	1	1350	1	1355	0	2398
gi 192970812 gb EU766597.1	98.67	1355	13	5	1	1350	1	1355	0	2398
gi 214023970 gb FJ36588.1	99.03	1336	13	0	1	1336	9	1344	0	2396
gi 192980961 gb EU774984.1	98.25	1368	24	0	1	1368	33	1400	0	2394
gi 219531267 gb FJ508072.1	98.88	1340	14	1	1	1340	16	1354	0	2390
gi 322145055 gb FJ159649.1	98.59	1351	18	1	1	1350	1	1351	0	2388
gi 214018819 gb FJ364432.1	98.45	1356	21	0	1	1356	1	1356	0	2388
gi 192968638 gb EU764423.1	98.53	1356	14	6	1	1350	1	1356	0	2388
gi 388930933 gb JQ185510.1	98.38	1362	10	12	1	1350	1	1362	0	2383
gi 192981465 gb EU775488.1	98.1	1368	26	0	1	1368	33	1400	0	2383
gi 110437046 gb DQ796646.1	98.1	1370	23	3	1	1368	33	1401	0	2383
gi 169278195 gb EU462720.1	98.03	1368	27	0	1	1368	21	1388	0	2377
gi 169278110 gb EU462635.1	98.03	1368	27	0	1	1368	21	1388	0	2377
gi 62765348 gb AY985858.1	98.31	1363	10	13	1	1350	1	1363	0	2377
gi 214026121 gb FJ371740.1	98.23	1356	24	0	1	1356	21	1376	0	2372
gi 169285018 gb EU469543.1	98.66	1339	17	1	1	1339	1	1338	0	2372
gi 214026227 gb FJ371846.1	98.51	1345	15	5	17	1356	1	1345	0	2368
gi 192970133 gb EU765918.1	98.3	1353	19	4	1	1350	1	1352	0	2368
gi 322145018 gb JF159612.1	98.3	1350	22	1	1	1350	1	1349	0	2364

gi 192970441 gb EU766226.1	98.16	1357	18	7	1	1350	1	1357	0	2361
gi 169278124 gb EU462649.1	99.54	1293	6	0	76	1368	1	1293	0	2355
gi 214018742 gb FJ364355.1	98.5	1332	20	0	1	1332	1	1332	0	2350
gi 388928549 gb JQ183666.1	97.87	1361	17	12	1	1350	1	1360	0	2342
gi 388934849 gb JQ189966.1	97.67	1375	7	25	1	1350	1	1375	0	2338
gi 319487794 gb HQ780655.1	97.45	1374	29	6	84	1453	65	1436	0	2338
gi 192981006 gb EU75029.1	97.51	1368	34	0	1	1368	33	1400	0	2338
gi 110446384 gb DQ806599.1	99.46	1285	7	0	84	1368	90	1374	0	2335
gi 110438803 gb DQ798403.1	99.38	1288	6	2	67	1352	15	1302	0	2333
gi 192969094 gb EU764879.1	97.78	1354	22	8	5	1350	6	1359	0	2327
gi 110440827 gb DQ800427.1	99.69	1272	4	0	97	1368	1	1272	0	2327
gi 110449954 gb DQ810169.1	97.37	1368	33	3	1	1368	25	1389	0	2324
gi 388928120 gb JQ183237.1	97.3	1368	17	20	1	1350	1	1366	0	2303
gi 110441294 gb DQ800894.1	97.01	1370	37	3	1	1368	33	1400	0	2300
gi 192970021 gb EU765806.1	99.45	1266	5	2	87	1350	93	1358	0	2298
gi 110436797 gb DQ796397.1	99.29	1261	8	1	109	1368	1	1261	0	2278
gi 169282930 gb EU467455.1	97.3	1335	35	1	1	1335	34	1367	0	2265
gi 61620028 gb AY850435.1	99.52	1243	6	0	67	1309	9	1251	0	2263
gi 169287428 gb EU471953.1	99.05	1261	9	3	111	1368	1	1261	0	2259
gi 169282916 gb EU467441.1	97.22	1331	36	1	1	1331	35	1364	0	2252
gi 169287473 gb EU471998.1	97.13	1288	32	3	84	1368	88	1373	0	2169
gi 169285037 gb EU469562.1	98.94	1130	12	0	222	1351	136	1265	0	2021
gi 110436563 gb DQ796163.1	97.39	1148	29	1	222	1368	235	1382	0	1953
gi 214017950 gb FJ363563.1	100	1049	0	0	307	1355	1	1049	0	1938
gi 84626925 gb DQ339838.1	100	1038	0	0	325	1362	1	1038	0	1917
gi 22324751 gb AF530344.1	99.81	1038	0	1	325	1362	1	1036	0	1905
gi 62753295 gb AY920178.1	99.42	1039	4	2	325	1362	1	1038	0	1884
gi 298391468 gb HM478330.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 298391173 gb HM478035.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 298390963 gb HM477825.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 298390562 gb HM477424.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 298390543 gb HM477405.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 298389976 gb HM476838.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 298389938 gb HM476800.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 298389589 gb HM476451.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 298389422 gb HM476284.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 298389379 gb HM476241.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 298389231 gb HM476093.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 298388837 gb HM475699.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258688844 emb FP083306.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 258688642 emb FP083302.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258688592 emb FP083252.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258687382 emb FP082637.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 258687350 emb FP082605.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 258687324 emb FP082579.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 258685368 emb FP081817.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258685243 emb FP075647.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258684705 emb FP075308.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258684256 emb FP075075.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258684063 emb FP081284.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258683838 emb FP081059.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 258681398 emb FP086361.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258681389 emb FP084854.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258680819 emb FP084484.1	100	1001	0	0	345	1345	1	1001	0	1849
gi 258680338 emb FP078738.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 258680294 emb FP078694.1	100	1001	0	0	344	1344	1	1001	0	1849
gi 298392754 gb HM479616.1	99.9	1001	1	0	345	1345	1	1001	0	1844
gi 298391479 gb HM478341.1	99.9	1001	1	0	344	1344	1	1001	0	1844
gi 298391366 gb HM478228.1	99.9	1001	1	0	344	1344	1	1001	0	1844
gi 298390465 gb HM477327.1	99.9	1001	1	0	345	1345	1	1001	0	1844
gi 298390259 gb HM477121.1	99.9	1001	1	0	345	1345	1	1001	0	1844
gi 298389656 gb HM476518.1	99.9	1001	1	0	345	1345	1	1001	0	1844
gi 298389217 gb HM476079.1	99.9	1001	1	0	345	1345	1	1001	0	1844
gi 258684984 emb FP081632.1	99.9	1001	1	0	344	1344	1	1001	0	1844
gi 258684844 emb FP075447.1	99.9	1001	1	0	345	1345	1	1001	0	1844
gi 258680857 emb FP084518.1	99.9	1001	1	0	344	1344	1	1001	0	1844
gi 258679890 emb FP078489.1	99.9	1001	1	0	345	1345	1	1001	0	1844
gi 258679841 emb FP078441.1	99.9	1001	1	0	344	1344	1	1001	0	1844

gi 258679682 emb FP078282.1	99.9	1001	1	0	345	1345	1	1001	0	1844		gi 310840826 gb GU105354.1	98.95	949	10	0	27	975	1	949	0	1698
gi 258685337 emb FP081786.1	99.8	1001	2	0	344	1344	1	1001	0	1838		gi 258680175 emb FP084224.1	97.3	1001	25	2	344	1344	1	999	0	1698
gi 258680032 emb FP078631.1	99.8	1001	2	0	344	1344	1	1001	0	1838		gi 310840972 gb GU105500.1	99.36	935	6	0	27	961	1	935	0	1694
gi 258679945 emb FP078544.1	99.8	1001	2	0	345	1345	1	1001	0	1838		gi 310841115 gb GU105643.1	98.94	944	10	0	32	975	2	945	0	1688
gi 258682949 emb FP080298.1	99.7	1001	3	0	344	1344	1	1001	0	1832		gi 73427009 gb DQ144095.1	99.57	925	4	0	511	1435	1	925	0	1687
gi 258681405 emb FP084870.1	99.7	1001	3	0	345	1345	1	1001	0	1832		gi 258681714 emb FP079519.1	97	1001	30	0	345	1345	1	1001	0	1683
gi 258681365 emb FP084830.1	99.7	1001	3	0	345	1345	1	1001	0	1832		gi 258689064 emb FP077679.1	97.01	1004	23	7	345	1345	1	1000	0	1681
gi 258680303 emb FP078703.1	99.7	1001	3	0	344	1344	1	1001	0	1832		gi 310840973 gb GU105501.1	99.04	936	9	0	27	962	1	936	0	1679
gi 258688156 emb FP083014.1	99.5	1001	5	0	344	1344	1	1001	0	1821		gi 310840928 gb GU105456.1	98.83	942	11	0	27	968	1	942	0	1679
gi 258687287 emb FP082542.1	99.1	1001	9	0	345	1345	1	1001	0	1799		gi 310840975 gb GU105503.1	98.93	936	10	0	27	962	1	936	0	1674
gi 258689133 emb FP077748.1	98.8	1001	12	0	344	1344	1	1001	0	1783		gi 310840927 gb GU105455.1	99.03	932	9	0	27	958	1	932	0	1672
gi 258679670 emb FP078270.1	98.8	1003	8	2	345	1345	1	1001	0	1783		gi 310840976 gb GU105504.1	98.82	936	11	0	27	962	1	936	0	1668
gi 62753249 gb AY920132.1	97.69	1039	21	2	325	1362	1	1037	0	1783		gi 310841112 gb GU105640.1	99.03	928	9	0	27	954	1	928	0	1664
gi 258689128 emb FP077743.1	98.6	1001	14	0	344	1344	1	1001	0	1772		gi 310841093 gb GU105621.1	97.72	967	20	2	27	992	1	966	0	1663
gi 258690090 emb FP078109.1	98.5	1001	15	0	345	1345	1	1001	0	1766		gi 310841111 gb GU105639.1	99.24	918	7	0	24	941	1	918	0	1657
gi 258679990 emb FP078589.1	98.5	1001	15	0	344	1344	1	1001	0	1766		gi 310841173 gb GU105701.1	99.34	912	4	2	28	938	1	911	0	1650
gi 258679817 emb FP078417.1	98.3	1001	17	0	344	1344	1	1001	0	1755		gi 310841037 gb GU105565.1	99.12	913	8	0	29	941	1	913	0	1642
gi 62753277 gb AY920160.1	97.02	1039	27	3	325	1362	1	1036	0	1744		gi 383385719 gb JQ799516.1	97.89	948	18	2	507	1453	1	947	0	1639
gi 310840978 gb GU105506.1	99.07	968	9	0	27	994	1	968	0	1738		gi 310840926 gb GU105454.1	99.12	910	8	0	27	936	1	910	0	1637
gi 258680809 emb FP084474.1	97.9	1002	20	1	344	1344	1	1002	0	1733		gi 310841163 gb GU105691.1	98.91	915	10	0	27	941	1	915	0	1635
gi 298392763 gb HM479625.1	97.8	1001	22	0	345	1345	1	1001	0	1727		gi 310840866 gb GU105394.1	99.01	912	9	0	27	938	1	912	0	1635
gi 258684128 emb FP074949.1	97.8	1001	22	0	345	1345	1	1001	0	1727		gi 310836892 gb GU104201.1	99.55	897	4	0	29	925	1	897	0	1635
gi 258688530 emb FP083190.1	97.7	1001	23	0	344	1344	1	1001	0	1722		gi 310840980 gb GU105508.1	99.33	901	6	0	27	927	1	901	0	1631
gi 258688471 emb FP083131.1	97.7	1001	23	0	344	1344	1	1001	0	1722		gi 310842043 gb GU106571.1	98.8	915	11	0	24	938	1	915	0	1629
gi 258687267 emb FP082522.1	97.7	1001	23	0	345	1345	1	1001	0	1722		gi 310841348 gb GU105876.1	99.12	905	8	0	24	928	1	905	0	1628
gi 258680259 emb FP078659.1	97.7	1002	21	2	345	1345	1	1001	0	1722		gi 310840979 gb GU105507.1	99.44	894	5	0	27	920	1	894	0	1624
gi 258679892 emb FP078491.1	97.7	1001	23	0	345	1345	1	1001	0	1722		gi 310842898 gb GU107426.1	98.9	908	10	0	31	938	1	908	0	1622
gi 258679706 emb FP078306.1	97.6	1002	22	2	344	1344	1	1001	0	1716		gi 310840856 gb GU105384.1	99.77	881	2	0	28	908	1	881	0	1616
gi 258687656 emb FP082713.1	97.5	1001	25	0	344	1344	1	1001	0	1711		gi 310840807 gb GU105335.1	99.77	880	2	0	26	905	1	880	0	1615
gi 310841110 gb GU105638.1	98.75	961	12	0	27	987	1	961	0	1709		gi 239620487 gb FJ651880.1	99.44	889	5	0	1	889	27	915	0	1615
gi 298392813 gb HM479675.1	97.4	1001	24	2	344	1344	1	999	0	1703		gi 310841256 gb GU105784.1	98.89	898	10	0	28	925	1	898	0	1604
gi 258684733 emb FP075336.1	97.4	1001	24	2	344	1344	1	999	0	1703		gi 239620493 gb FJ651886.1	99.21	889	7	0	1	889	27	915	0	1604
gi 258680334 emb FP078734.1	97.3	1001	27	0	344	1344	1	1001	0	1700		gi 310842899 gb GU107427.1	98.36	912	15	0	27	938	1	912	0	1602
gi 258680314 emb FP078714.1	97.3	1001	27	0	345	1345	1	1001	0	1700		gi 310842632 gb GU107160.1	98.78	900	11	0	27	926	1	900	0	1602

gi 310838465 gb GU102993.1	98.89	897	10	0	24	920	1	897	0	1602
gi 310842230 gb GU106758.1	98.88	896	10	0	24	919	1	896	0	1600
gi 310843290 gb GU107818.1	99.77	868	2	0	24	891	1	868	0	1592
gi 310842840 gb GU107368.1	98.45	905	13	1	28	931	1	905	0	1592
gi 310841113 gb GU105641.1	98.55	898	13	0	29	926	1	898	0	1587
gi 291331103 gb GU957635.1	98.88	889	10	0	1	889	27	915	0	1587
gi 310842044 gb GU106572.1	98.66	894	12	0	24	917	1	894	0	1585
gi 310841152 gb GU105680.1	98.77	892	10	1	27	918	1	891	0	1585
gi 310840857 gb GU105385.1	98.55	897	13	0	28	924	1	897	0	1585
gi 291328328 gb GU954860.1	99.43	873	5	0	1	873	29	901	0	1585
gi 310841879 gb GU106407.1	99.54	868	4	0	33	900	2	869	0	1581
gi 295651187 gb HM013421.1	98.76	889	11	0	1	889	28	916	0	1581
gi 239620005 gb F4651398.1	98.76	889	11	0	1	889	16	904	0	1581
gi 291328488 gb GU955020.1	98.76	889	10	1	2	889	3	891	0	1580
gi 310841909 gb GU106437.1	99.54	866	4	0	27	892	1	866	0	1578
gi 239619923 gb F4651316.1	99.42	869	5	0	21	889	1	869	0	1578
gi 169279765 gb EU464290.1	98.02	908	18	0	461	1368	18	925	0	1578
gi 291331104 gb GU957636.1	98.65	889	12	0	1	889	27	915	0	1576
gi 239620484 gb F4651877.1	98.65	889	12	0	1	889	27	915	0	1576
gi 239620008 gb F4651401.1	98.65	889	12	0	1	889	17	905	0	1576
gi 310839154 gb GU103682.1	98.98	879	9	0	27	905	1	879	0	1574
gi 310841445 gb GU105973.1	99.77	857	2	0	21	877	1	857	0	1572
gi 291328286 gb GU954818.1	98.54	889	13	0	1	889	28	916	0	1570
gi 239620488 gb F4651881.1	98.54	889	13	0	1	889	27	915	0	1570
gi 239619382 gb F4650775.1	98.54	889	13	0	1	889	27	915	0	1570
gi 310840977 gb GU105505.1	99.42	864	5	0	27	890	1	864	0	1568
gi 291328529 gb GU955061.1	98.43	889	14	0	1	889	28	916	0	1565
gi 310842046 gb GU106574.1	98.53	885	13	0	24	908	1	885	0	1563
gi 310841342 gb GU105870.1	99.77	852	2	0	24	875	1	852	0	1563
gi 310838205 gb GU102733.1	98.32	892	13	2	25	916	1	890	0	1563
gi 310837149 gb GU101677.1	99.42	860	5	0	33	892	3	862	0	1561
gi 310835970 gb GU100498.1	99.3	863	6	0	29	891	1	863	0	1561
gi 291331110 gb GU957642.1	98.31	889	15	0	1	889	27	915	0	1559
gi 239619213 gb F4650606.1	98.31	888	15	0	1	888	27	914	0	1557
gi 310842067 gb GU106595.1	99.53	854	4	0	24	877	1	854	0	1555
gi 310841114 gb GU105642.1	99.3	860	6	0	29	888	1	860	0	1555
gi 239619393 gb F4650786.1	98.63	877	12	0	1	877	27	903	0	1554
gi 529080432 gb KF229853.1	98.96	866	9	0	24	889	1	866	0	1550
gi 310840808 gb GU105336.1	98.74	872	11	0	27	898	1	872	0	1550
gi 239619345 gb F4650738.1	98.63	874	12	0	1	874	27	900	0	1548
gi 310841380 gb GU105908.1	99.53	849	4	0	29	877	1	849	0	1546
gi 291331651 gb GU958183.1	97.98	890	16	2	1	889	28	916	0	1543
gi 310838473 gb GU103001.1	99.64	843	3	0	24	866	1	843	0	1541
gi 310839536 gb GU104064.1	98.62	870	10	2	24	892	1	869	0	1539
gi 291331102 gb GU957634.1	99.41	848	5	0	42	889	1	848	0	1539
gi 239620288 gb F4651681.1	98.73	866	11	0	1	866	11	876	0	1539
gi 310838146 gb GU102674.1	98.73	866	10	1	25	890	1	865	0	1537
gi 5290803001 gb KF234222.1	98.62	867	12	0	1	867	23	889	0	1535
gi 529080529 gb KF229950.1	98.84	861	10	0	2	862	1	861	0	1535
gi 291329410 gb GU955942.1	98.73	864	11	0	1	864	14	877	0	1535
gi 529080654 gb KF230075.1	99.06	854	8	0	1	854	21	874	0	1533
gi 310841035 gb GU105631.1	98.5	868	13	0	28	895	1	868	0	1531
gi 169287659 gb EU472184.1	97.44	898	23	0	471	1368	1	898	0	1531
gi 310838633 gb GU103161.1	99.64	837	3	0	33	869	3	839	0	1530
gi 310842151 gb GU106679.1	99.4	840	5	0	27	866	1	840	0	1524
gi 310842099 gb GU106627.1	99.4	840	5	0	27	866	1	840	0	1524
gi 310841955 gb GU106483.1	99.4	840	5	0	24	863	1	840	0	1524
gi 310841878 gb GU106406.1	99.4	840	5	0	24	863	1	840	0	1524
gi 310838817 gb GU103345.1	99.64	834	3	0	29	862	1	834	0	1524
gi 310838576 gb GU103104.1	99.52	837	3	1	27	863	1	836	0	1522
gi 310841036 gb GU105564.1	98.38	865	14	0	28	892	1	865	0	1520
gi 164454293 obj AB265290.1	97.53	889	21	1	478	1366	1	888	0	1519
gi 310841954 gb GU106482.1	99.05	845	8	0	33	877	1	845	0	1517
gi 310841907 gb GU106435.1	99.4	835	5	0	29	863	1	835	0	1515
gi 310838717 gb GU103245.1	99.52	833	3	1	33	865	3	834	0	1515
gi 310833518 gb GU098046.1	99.52	832	4	0	33	864	1	832	0	1515
gi 310841908 gb GU106436.1	99.28	837	6	0	27	863	1	837	0	1513
gi 118136060 gb EF071471.1	98.82	849	10	0	1	849	20	868	0	1513

gi 310841536 gb GU106064.1	99.16	838	7	0	29	866	1	838	0	1509
gi 310841078 gb GU105606.1	99.17	839	6	1	27	864	1	839	0	1509
gi 310838778 gb GU103306.1	99.16	838	7	0	28	865	1	838	0	1509
gi 310841959 gb GU106487.1	99.28	835	5	1	29	863	1	834	0	1507
gi 310841378 gb GU105906.1	99.05	840	8	0	24	863	1	840	0	1507
gi 310842517 gb GU107045.1	98.15	863	16	0	27	889	1	863	0	1506
gi 310833629 gb GU098157.1	98.82	845	10	0	22	866	1	845	0	1506
gi 310840786 gb GU105314.1	98.93	840	9	0	27	866	1	840	0	1502
gi 310839537 gb GU104065.1	98.81	843	10	0	24	866	1	843	0	1502
gi 310841875 gb GU106403.1	98.47	851	13	0	27	877	1	851	0	1500
gi 310840855 gb GU105383.1	98.7	845	11	0	27	871	1	845	0	1500
gi 310837024 gb GU101552.1	98.36	856	11	3	27	879	1	856	0	1500
gi 310838419 gb GU102947.1	99.88	814	1	0	50	863	4	817	0	1498
gi 110440903 gb DQ800503.1	99.04	835	8	0	1	835	33	867	0	1498
gi 310842583 gb GU107111.1	98.36	852	14	0	25	876	1	852	0	1496
gi 310840877 gb GU105405.1	98.81	840	10	0	27	866	1	840	0	1496
gi 310837892 gb GU102420.1	98.81	840	10	0	24	863	1	840	0	1496
gi 310833517 gb GU098045.1	99.16	831	7	0	33	863	1	831	0	1496
gi 310842069 gb GU106596.1	98.24	854	15	0	24	877	1	854	0	1495
gi 399141153 gb JQ265125.1	98.69	842	10	1	38	878	1	842	0	1493
gi 310842262 gb GU106790.1	99.39	823	5	0	21	843	1	823	0	1493
gi 310833423 gb GU097951.1	99.16	829	7	0	36	864	4	832	0	1493
gi 310842231 gb GU106759.1	98.58	843	12	0	24	866	1	843	0	1491
gi 310842154 gb GU106682.1	98.69	840	11	0	24	863	1	840	0	1491
gi 310842135 gb GU106663.1	98.69	840	11	0	27	866	1	840	0	1491
gi 310842045 gb GU106573.1	98.69	840	11	0	24	863	1	840	0	1491
gi 310841956 gb GU106484.1	98.69	840	11	0	24	863	1	840	0	1491
gi 310842138 gb GU106666.1	98.57	840	12	0	27	866	1	840	0	1485
gi 310838495 gb GU103023.1	99.03	828	8	0	24	851	1	828	0	1485
gi 239620227 gb FJ651620.1	98.34	844	14	0	35	878	2	845	0	1482
gi 399141562 gb JQ265534.1	98.68	835	9	2	1	833	65	899	0	1480
gi 310842150 gb GU106678.1	98.57	837	12	0	27	863	1	837	0	1480
gi 310842101 gb GU106629.1	98.45	840	13	0	24	863	1	840	0	1480
gi 310842097 gb GU106625.1	98.45	840	13	0	27	866	1	840	0	1480

gi 310841108 gb GU105636.1	97.79	861	14	5	39	895	2	861	0	1480
gi 310837285 gb GU101813.1	98.91	828	9	0	24	851	1	828	0	1480

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